



The Advantages of Mesenchymal Stem Cell-Derived Extracellular Vesicles as Drug Delivery Systems

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Abstract

Extracellular vesicles (EVs) are nanosized particles, secreted by most, if not all cell types, enclosed by a bilayer phospholipid membrane, and mediate crucial intercellular communications by carrying and transporting many active biological molecules. These naturally occurred nanoparticles are emerging as a promising drug delivery system. In particular, EVs from mesenchymal stem cells (MSC-EVs) offer numerous advantages as drug delivery vehicles due to their unique features, as briefly reviewed below in Figure 1.

REVIEW SUMMARY

The advantages of mesenchymal stem cell-derived extracellular vesicles as drug delivery systems

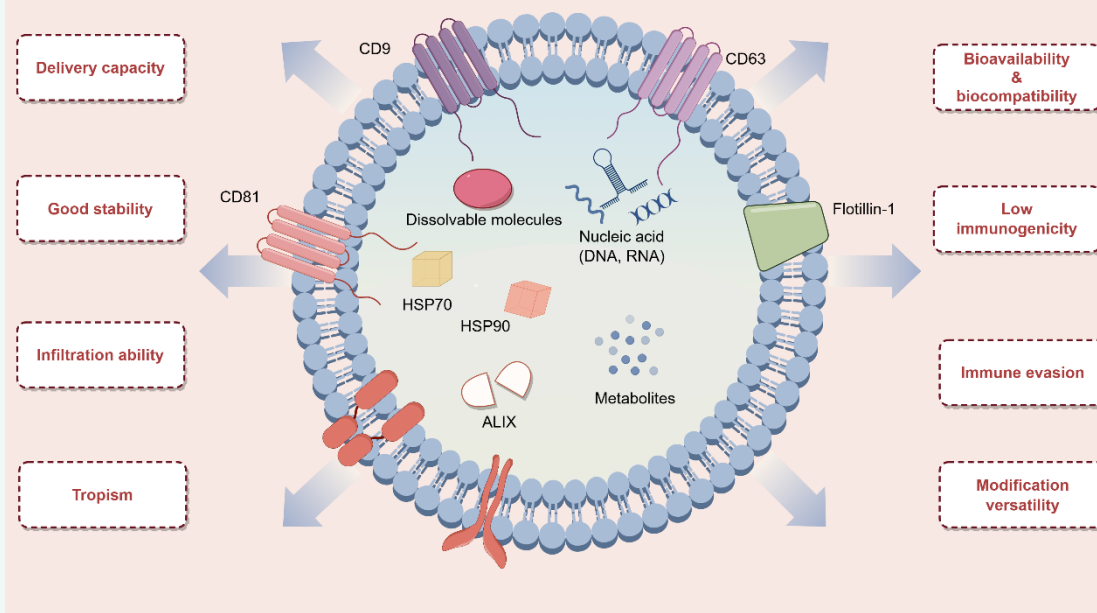


Figure 1: Summary of the Advantages of Mesenchymal Stem Cell-Derived Extracellular Vesicles as Drug Delivery Systems.

Submitted: 30 June, 2025 | Accepted: 08 August, 2025 | Published: 09 August, 2025

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Citation: Kuang X, Peng J, Peng Y, Huang C, Yuan Z (2025) The Advantages of Mesenchymal Stem Cell-Derived Extracellular Vesicles as Drug Delivery Systems. SM J Biol 7: 4.

THE CAPACITY TO DELIVER BOTH SOLUBLE CARGOES AND MEMBRANE BOUND THERAPEUTIC MOLECULES

MSC-EVs have the capability to deliver a variety of therapeutic cargoes, including soluble agents like small molecule drugs and RNAs as well as membrane-integrated therapeutic molecules such as proteins and antibodies [1,2]. By encapsulating these therapeutic molecules, MSC-EVs not only enhance the stability and bioavailability of the drugs but also enable co-delivery of synergistic drugs [3,4]. For example, chemotherapy drugs doxorubicin with miR-159 or 5-fluorouracil (5-FU) with miR-21i could be co-delivered for synergistic anticancer therapies [5,6]. Previously we genetically transduced MSCs to express the proapoptotic tumor necrosis factor-related apoptosis-inducing ligand (TRAIL) [7] and found the transduced cells secreted EV-membrane incorporated TRAIL



(EV-T) [8]. Further we encapsulated the chemotherapy dinaciclib (Dina) into EV-T to fabricate a complexed nanodrug Dina@EV-T, which overcame TRAIL resistance and showed strikingly augmented anticancer efficacy by either intravenous (i.v.) [4] or nebulized administration [9]. Additionally, the CDK9-targeting siRNA could be loaded into EV-T for precision anticancer medicine [10]. Therefore, MSC-EV mediated co-delivery of drugs enables high efficacy of combinatory therapies.

GOOD STABILITY BOTH *IN VITRO* AND *IN VIVO*

As the natural drug carrier, MSC-EVs have shown good stability both *in vitro* and *in vivo* [9]. The membrane structure of EVs forms a natural and biocompatible platform to not only protect the enclosed drugs from premature degradation by enzymes and other degradative processes, but also help in maintaining the stability of labile therapeutics. The *in vivo* stability of EVs was proposed to be further enhanced by the coating of an albumin-enriched protein corona and consequent immune evasion [11].

THE ABILITY TO INFILTRATE AND PENETRATE TISSUES AND THE BLOOD-BRAIN BARRIER

EVs have innate capacity to cross various biological barriers including the blood-brain barrier (BBB) [12]. The tissue permeability of nanomedicines was traditionally assumed to be mainly mediated by the enhanced permeability and retention (EPR) effect. However, the advancements in this area have unveiled a possible transcytosis mechanism, by which EVs can migrate through endothelial cells [13]. This property highlights the attractive potential of EV-based therapies for brain disorders and tumors [14]. Indeed, MSC-EVs have been recently harnessed to successfully deliver therapeutic siRNAs to the striatum of mice brain, leading to the synergistic alleviation of neuronal death in a model of Parkinson's disease (PD) [12].

TISSUE AND ORGAN TROPISMS

The tropism of MSC-EVs to certain damaged, inflammatory or diseased tissue and organs enable targeted delivery of therapeutic agents [4-15]. This phenomenon is likely mediated by EV surface molecules, such as tetraspanins, latex adhesion proteins, and integrins [16]. As an instance, MSC-EVs showed preferential tropism to animal acute lung injury (ALI), by contrast, HEK293T cells-derived EVs were mainly accumulated in the spleen and liver [17]. The systemically infused MSC-EVs were found to penetrate and accumulate in tumors, suggesting their feasibility for delivery of tumor-targeting therapy. [4] For example, doxorubicin encapsulated MSC-EVs were revealed to home to osteosarcoma via a CXCR4-SDF1 axis, resulting in enhanced anticancer activity [18].

GOOD BIOAVAILABILITY AND BIOCOMPATIBILITY

Compared to synthetic carriers such as liposomes or other nanoparticles, MSC-EVs demonstrated good biocompatibility and bioavailability as drug delivery systems [1,2]. As carriers, EVs can transport their cargo across cell membranes to specific intracellular locations. Importantly, the lipid membrane structure renders MSC-EVs good biological barrier penetrating capacity, *in vivo* stability, reduced immune clearance, and consequently improved drug delivery efficiency [19]. Furthermore, the low immunogenicity and good tolerability of MSC-EVs allow for their safe use *in vivo*, minimizing the risk of eliciting adverse reactions [4]. The desirable biocompatibility and bioavailability make MSC-EVs ideal candidates for drug delivery carriers.

LOW IMMUNOGENICITY AND GOOD SAFETY

MSC-EVs are increasingly recognized for their low immunogenicity, a characteristic pivotal to their therapeutic potential in various diseases [20]. Certain EV surface components, such as galectins, integrins, and tetraspanins, play a masking role from the immune system leading to the low immunogenicity of MSC-EVs [21]. Additionally, the EV surface glycans and lipids also act as key signaling molecules to influence the immunogenicity, with the glycan composition dictating cellular internalization and biodistribution, while lipids contributing to intercellular communication and immune modulation [21]. It is well-known that major histocompatibility complex (MHC) molecules are crucial for antigen presentation to trigger immune responses, thus play an important role for immunogenicity. MSC-EVs have been revealed to be negative for MHC expression, indicating their low immunogenicity [22]. Actually, the systemic administration of MSC-EVs did not cause any adverse side effects on liver function, blood cell counting and organ physiology in experimental animals [4].

In the realm of safety evaluation, the administration of extracellular vesicles (EVs) has demonstrated notable superiority. Preclinical studies have revealed the superior efficacy, safety, and versatility of MSC-EV therapies compared to the MSC therapy, suggesting the satisfactory safety of MSC-EV delivery of therapeutics for disease treatment [23]. Indeed, over 20 completed or ongoing MSC-EV clinical trials have demonstrated good safety and certain efficacies in various diseases.[20] Moreover, in a phase 1 clinical trial, 24 healthy volunteers were administered of $2-16 \times 10^8$ allogeneic MSC-EV particles by inhalation, and all showed good tolerance to the infusion without any adverse reactions observed [24].

IMMUNE EVASION

The application of nanomedicine faces a huge challenge, i.e., the rapid uptake and subsequent clearance of nanoparticles from the bloodstream by the mononuclear phagocyte system (MPS) [25]. Interestingly, the integrin-associated protein CD47 was found to express on MSC-EVs [4] and act as a marker of self to prevent clearance by the MPS in the liver and spleen [26]. This feature allows phagocytic evasion of MSC-EVs, and provides an approach for improving pharmacokinetics of therapeutics and thus potentially enhancing therapeutic efficacies. This advantage underscores the versatility and potential of MSC-EVs as a drug delivery vehicle for therapeutic applications [27].

MODIFICATION FLEXIBILITY

The enclosed lipid membrane structure and specific surface compositions enable flexible modification of MSC-EVs for either the targeted delivery of therapeutics or EV labelling and tracking within cells or tissues. There are various EV surface modification strategies available, including genetic engineering or metabolic engineering of EV-producing cells, click chemistry, ligand-receptor interaction, hydrophobic interaction, and anchoring peptide or aptamer-mediated modifications. [28] Also, different labelling approaches have been developed for EV imaging, pharmacokinetic investigation, or examination of *in vivo* biodistribution, such as bioluminescent, fluorescent, or radioactive labelling. [4-28] Furthermore, both drug pre-loading and post-loading strategies can be applied to engineer MSC-EVs for enhanced therapeutic efficacies. [4-30] Despite the aforementioned versatility and potential, one must be careful to determine MSC-EV modification strategies, considering the modifications may cause undesired immunogenicity, reduced physicochemical stability,

and the biosafety concern.[28]

In summary (Figure.2), MSC-derived extracellular vesicles offer several advantages as drug delivery systems. They exhibit high biocompatibility and low immunogenicity, ensuring safe administration. These vesicles can efficiently target specific tissues and cells, enhancing therapeutic efficacy. Additionally, they are capable of carrying a diverse range of therapeutic molecules, including proteins, RNA, and small molecules. Their nanoscale size allows for easy penetration into tissues, and they can be readily produced and stored, making them a practical option for clinical applications.

ACKNOWLEDGEMENTS

Z.Q.Y. is a China Talented Scholar Scheme Research Fellow in the School of Biomedical and Pharmaceutical Sciences and is supported by the Guangzhou Provincial Talented Scholar Foundation (220418137).

AUTHOR CONTRIBUTIONS

Z.Q.Y provided the theme and framework for the review, and along with C.H, he was responsible for the final proofreading and editing of the article, as well as the approval of the final version. X.B.K and J.P contributed equally to the drafting and revision, and Y.Y.P participated the writing of the manuscript.

FUNDING

This work has been supported by National Natural Science Foundation of China (grant number 82173850), Guangdong Medical Science and Technology Research Fund (A2024625), and Guangzhou Science and Technology Planning Project (202103000002).

CONFLICT OF INTERESTS

The authors declare no any financial, proprietary, or commercial interests in the writing of this article.

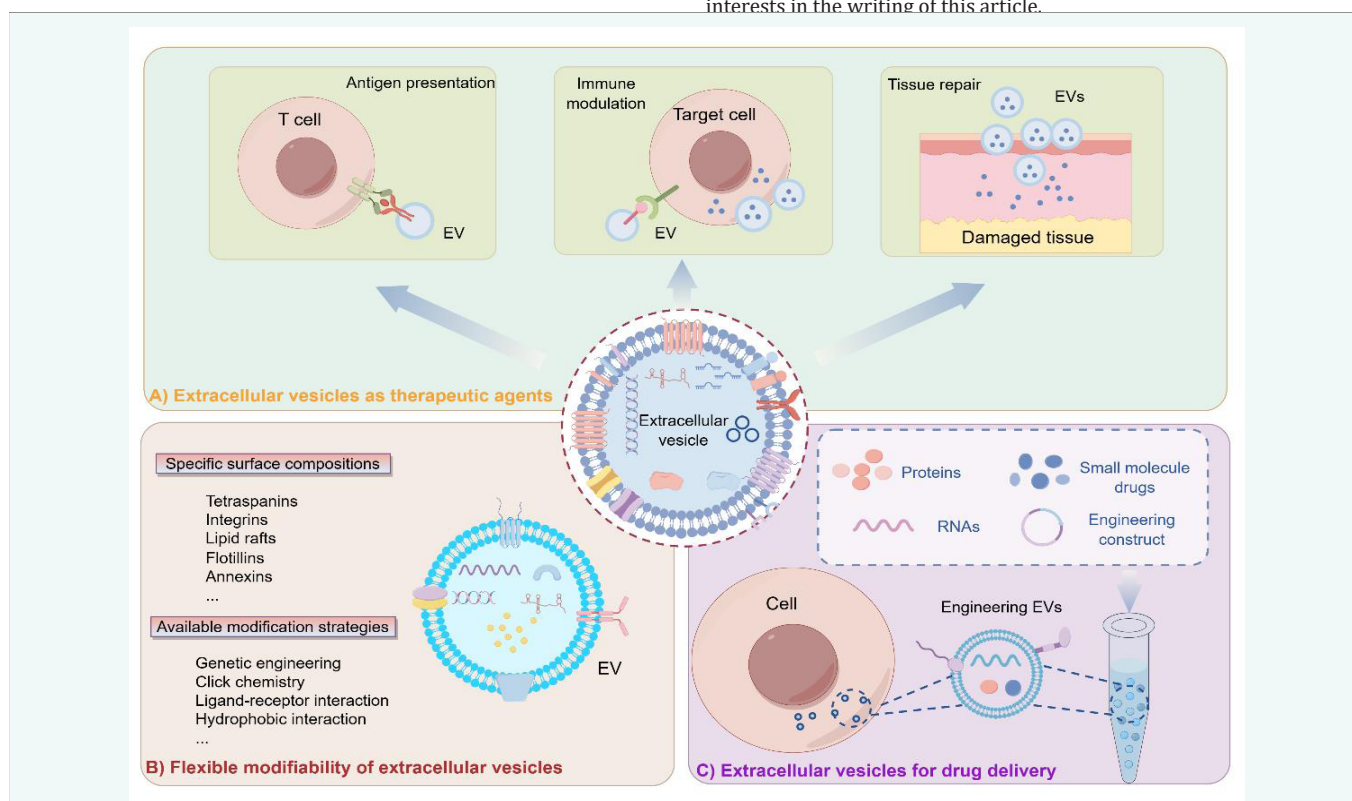


Figure 2: Extracellular vesicles mediate antigen presentation, immune modulation, and tissue repair. They can also be used as adaptable drug delivery vehicles and diagnostic tools.

REFERENCES

1. Liu Q, Li D, Pan X, Liang Y. Targeted therapy using engineered extracellular vesicles: principles and strategies for membrane modification. *J Nanobiotechnology*. 2023; 21: 334.
2. Rezaie J, Nejati V, Mahmoodi M, Ahmadi M. Mesenchymal stem cells derived extracellular vesicles: A promising nanomedicine for drug delivery system. *Biochem Pharmacol*. 2022; 203: 115167.
3. Chen H, Yao H, Chi J, Li C, Liu Y, Yang J et al. Engineered exosomes as drug and RNA co-delivery system: new hope for enhanced therapeutics? *Front Bioeng Biotechnol*. 2023; 11: 1254356.
4. Ke C, Hou H, Su K, Huang C, Yuan Q, Li S, et al. Extracellular vesicle-mediated co-delivery of TRAIL and dinaciclib for targeted therapy of resistant tumors. *Biomater Sci*. 2022; 10: 1498-1514.
5. Gong C, Tian J, Wang Z, Gao Y, Wu X, Ding X, et al. Functional exosome-mediated co-delivery of doxorubicin and hydrophobically modified microRNA 159 for triple-negative breast cancer therapy. *J Nanobiotechnology*. 2019; 17: 93.
6. Liang G, Zhu Y, Ali DJ, Tian T, Xu H, Si K et al. Engineered exosomes for targeted co-delivery of miR-21 inhibitor and chemotherapeutics to reverse drug resistance in colon cancer. *J Nanobiotechnology*. 2020; 18: 10.



7. Yuan Z, Kolluri KK, Sage EK, Gowers KH, Janes SM. Mesenchymal stromal cell delivery of full-length tumor necrosis factor-related apoptosis-inducing ligand is superior to soluble type for cancer therapy. *Cytotherapy*. 2015; 17: 885-896.
8. Yuan Z, Kolluri KK, Gowers KH, Janes SM. TRAIL delivery by MSC-derived extracellular vesicles is an effective anticancer therapy. *J Extracell Vesicles*. 2017; 6: 1265291.
9. Yuan Q, Su K, Li S, Long X, Liu L, Yang M et al. Pulmonary Delivery of Extracellular Vesicle-Encapsulated Dinaciclib as an Effective Lung Cancer Therapy. *Cancers (Basel)*. 2022; 14: 3550.
10. Yuan Q, Su K, Li S, Long X, Liu L, Sun J et al. Selective CDK9 knockdown sensitizes TRAIL response by suppression of antiapoptotic factors and NF-kappaB pathway. *Apoptosis*. 2023; 28: 1060-1075.
11. Liam-Or R, Faruqu FN, Walters A, Han S, Xu L, Wang JT et al. Cellular uptake and in vivo distribution of mesenchymal-stem-cell-derived extracellular vesicles are protein corona dependent. *Nat Nanotechnol*. 2024; 19: 846-855.
12. Geng Y, Long X, Zhang Y, Wang Y, You G, Guo W et al. FTO-targeted siRNA delivery by MSC-derived exosomes synergistically alleviates dopaminergic neuronal death in Parkinson's disease via m6A-dependent regulation of ATM mRNA. *J Transl Med*. 2023; 21: 652.
13. Ramos-Zaldívar HM, Polakovicova I, Salas-Huenuleo E, Corvalán AH, Kogan MJ, Yefi CP et al. Extracellular vesicles through the blood-brain barrier: a review. *Fluids Barriers CNS*. 2022; 19: 60.
14. Bhom N, Somandi K, Ramburrun P, Choonara YE. Extracellular nanovesicles as neurotherapeutics for central nervous system disorders. *Expert Opin Drug Deliv*. 2025; 22: 69-84.
15. Kalluri R, LeBleu VS. The biology, function, and biomedical applications of exosomes. *Science*. 2020; 367.
16. Edelmann MJ, Kima PE. Current understanding of extracellular vesicle homing/tropism. *Zoonoses (Burlingt)* 2022; 2: 14.
17. Tieu A, Stewart DJ, Chwastek D, Lansdell C, Burger D, Lalu MM. Biodistribution of mesenchymal stromal cell-derived extracellular vesicles administered during acute lung injury. *Stem Cell Res Ther*. 2023; 14: 250.
18. Wei H, Chen F, Chen J, Lin H, Wang S, Wang Y et al. Mesenchymal Stem Cell Derived Exosomes as Nanodrug Carrier of Doxorubicin for Targeted Osteosarcoma Therapy via SDF1-CXCR4 Axis. *Int J Nanomedicine*. 2022; 17: 3483-3495.
19. Xu F, Luo S, Lu P, Cai C, Li W, Li C. Composition, functions, and applications of exosomal membrane proteins. *Front Immunol*. 2024; 15: 1408415.
20. Lotfy A, AboQuella NM, Wang H. Mesenchymal stromal/stem cell (MSC)-derived exosomes in clinical trials. *Stem Cell Res Ther*. 2023; 14: 66.
21. Xia Y, Zhang J, Liu G, Wolfram J. Immunogenicity of Extracellular Vesicles. *Adv Mater*. 2024; 36: e2403199.
22. Desgeorges A, Hollerweger J, Lassacher T, Rohde E, Helmbrecht C, Gimona M. Differential fluorescence nanoparticle tracking analysis for enumeration of the extracellular vesicle content in mixed particulate solutions. *Methods*. 2020; 177: 67-73.
23. Vizoso FJ, Eiro N, Cid S, Schneider J, Perez-Fernandez R. Mesenchymal Stem Cell Secretome: Toward Cell-Free Therapeutic Strategies in Regenerative Medicine. *Int J Mol Sci*. 2017; 18: 1852.
24. Shi MM, Yang QY, Monsel A, Yan JY, Dai CX, Zhao JY et al. Preclinical efficacy and clinical safety of clinical-grade nebulized allogenic adipose mesenchymal stromal cells-derived extracellular vesicles. *J Extracell Vesicles*. 2021; 10: e12134.
25. Zelepukin IV, Shevchenko KG, Deyev SM. Rediscovery of mononuclear phagocyte system blockade for nanoparticle drug delivery. *Nat Commun*. 2024; 15: 4366.
26. Creeden JF, Sevier J, Zhang JT, Lapitsky Y, Brunicardi FC, Jin G et al. Smart exosomes enhance PDAC targeted therapy. *J Control Release*. 2024; 368: 413-429.
27. Ulpiano C, da Silva CL, Monteiro GA. Bioengineered Mesenchymal-Stromal-Cell-Derived Extracellular Vesicles as an Improved Drug Delivery System: Methods and Applications. *Biomedicines*. 2023; 11: 1231.
28. Salunkhe S, Dheeraj, Basak M, Chitkara D, Mittal A. Surface functionalization of exosomes for target-specific delivery and in vivo imaging & tracking: Strategies and significance. *J Control Release*. 2020; 326: 599-614.
29. Walker S, Busatto S, Pham A, Tian M, Suh A, Carson K et al. Extracellular vesicle-based drug delivery systems for cancer treatment. *Theranostics*. 2019; 9: 8001-8017.
30. Pascucci L, Coccè V, Bonomi A, Ami D, Ceccarelli P, Ciusani E et al. Paclitaxel is incorporated by mesenchymal stromal cells and released in exosomes that inhibit in vitro tumor growth: a new approach for drug delivery. *J Control Release*. 2014; 192: 262-270.