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*Corresponding author

Heba Elmansi, Analytical chemistry department, Faculty of Pharmacy, University of Mansoura, Egypt, Email: dr_heba85@hotmail.com

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Keywords CTZ: Cetirizine; co-formulated tablet; HPLC; SAL: Salbutamol

Research Article

A Validated Reversed Phase HPLC Method for Simultaneous Determination of the antihistaminic Cetirizine and Beta2-adrenergic agonist Salbutamol in their Co-formulated Tablets

Fatma Ahmed Aly, Nahed EL-Enany, Heba Elmansi* and Amany Nabil

Department of Analytical Chemistry, Faculty of Pharmacy, Mansoura University, Egypt

Abstract

New HPLC method is adopted in this research for simultaneous determination of Cetirizine (CTZ) and salbutamol (SAL) in their tablets. The developed method used C18 column and a mobile phase composed of methanol: 0.1M phosphate buffer in the ratio (80:20 v/v) operating at pH 3.5.Nimsulide was used as internal standard (IS).The peak area ratio - concentration plot indicated the linearity over ranges of 5-50 and 4-80 μ g/mL for CTZ and SAL with limit of detection of 0.57 and 2.00 μ g/mL respectively. Comparing the proposed method with a comparison method revealed that there was no significant difference between the two methods in regards to accuracy and precision.

Introduction

Cetirizine (CTZ) and Salbutamol (SAL) are co-formulated for treatment of common cold and allergy [1]. Cetirizine (CTZ, Figure 1a) is [2-[4-[(4-Chlorophenyl) phenylmethyl]-1-piperazinyl] ethoxy] acetic acid). It is long acting antihistaminic which used for relief of allergic conditions [2]. The BP described non-aqueous potentiometric titration method for CTZ determination [3]. Previously published methods described for determinations of CTZ include HPLC either in pharmaceutical preparations [4-6] or in biological fluids [7,8], fluorimetry [9] and spectrophotometry [10].

Salbutamol (SAL, Figure 1b) has a chemical name of 2-tert-Butylamino-1-(4-hydroxy-3-hydroxymethylphenyl) ethanol. It is B2-receptor agonist which acquiring a bronchodilation action and used for the treatment of chronic obstructive pulmonary disease and in managing asthma [2]. The BP recommended non-aqueous potentiometric titration method using perchloric acid as a titrant [3]. Determination of salbutamol was reported in several former publications such as HPLC [11-13], spectrophotometry [14,15], flow injection [16,17], capillary electrophoresis [18].

The combination of CTZ and SAL with a pharmaceutical ratio of 5:2 respectively is present under the trade name Vetirex1. This combination is used for common cold and allergic conditions. Up till now; no reported method was described for the determination of CTZ and SAL. This encourages us to develop a simple HPLC method for their simultaneous determination.

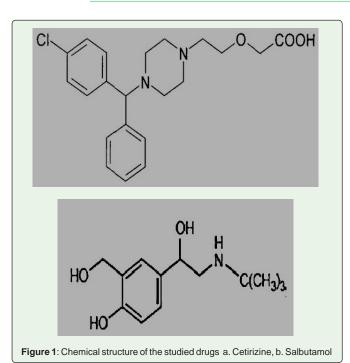
Experimental

HPLC analysis was performed using Perkin Elmer TM series 200 chromatograph (USA) with 200 μ L loop supplied with injector valve of Rheodyne. Series 200 UV/VIS. Detector was used and set at wave length 230 nm. The column used for analysis was Shimadzu VP-ODS column (250 mm x 4.6 mm i.d., 5 μ m particle size).

Materials and reagents

Cetirizine (batch # 3003CZ8RJ) pure sample was provided from Apex Co. (Cairo, Egypt). Its percentage purity was found to be 99.95(as labeled). Salbutamol (batch # 511/55/03/5018) was obtained from Pharaonia Co. Alex, Egypt. Its purity was found to be 100.15% as labeled from the manufacturer. Nimesulide (IS) was of 99.90% purity, with batch No# 000604. It is provided from Pharaonia Co., Alex, Egypt. Zyrtec* tablet (batch #6221045001829) labeled to contain 10 mg CTZ/tablet, manufactured by Glaxosmithkline Company obtained from a community pharmacy. Ventolin* (2mg) tablet (batch #6221045000969) contains 2 mg SAL per tablet, produced by Glaxosmithkline Company bought from a community pharmacy. Solium dihydrogen phosphate and sodium hydroxide were provided from ADWIC CO., Egypt. Solvents (HPLC grade) were obtained from Sigma-Aldrich (Germany). O-phosphoric acid (85 % w/v) was provided from Riedel-deHaen (Germany).

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Optimum chromatographic conditions

Shimadzu VP-ODS column (250 mm x 4.6 mm i.d., 5 μ m particle size), Japan was used as a chromatographic column. The separation was carried out at room temperature at 1 mL/min flow rate. The mobile phase composed of methanol: 0.1M sodium dihydrogen phosphate (80:20 v/v). PH was justified to 3.5. The detector wavelength was set at 230nm.

Standard solutions

Accurately weighed 20 mg of CTZ, SAL and IS drug was dissolved in 100 mL of methanol to obtain stock solutions of 200 $\mu g/mL$.

General procedures

Construction of calibration graphs

Aliquot volumes of a stock solution of CTZ and SAL were taken into a set of 10 mL volumetric flask. The solutions were diluted to the volume with the mobile phase at pH 3.5 to obtain a final concentration of 5-50 and 4-80 μ g/mL for CTZ and SAL, respectively. NIM (IS) in 20 μ g/mL concentration was added. Plotting peak area ratio versus the concentration in μ g/mL gave us the corresponding calibration graphs.

Analysis of laboratory- prepared mixture of CTZ and SAL

Laboratory prepared mixture of CTZ and SAL were freshly prepared by mixing different volumes of the drug in the ratio5:2, respectively together with the IS and then the procedure for chromatographic analysis under optimum conditions were performed. The mean percent recoveries of each drug were determined from the previously plotted calibration curve.

Analysis of laboratory- prepared co-formulated tablets

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Aliquots of the accurately weighed powdered tablets equivalent to 5.0 mg CTZ and 2.0 mg SAL were transferred to volumetric flasks 100 mL volume. Nearly 80 mL methanol was added and undergoes sonication for 10 minutes. The volume was then completed with methanol and filtered. Aliquots of this solution with the IS were transferred to a series of 10 mL volumetric flasks to obtain suitable concentrations within the working range. The mobile phase containing the mixture was eluted under suitable chromatographic conditions. The mean percent content was calculated using the regression equations or calibration graphs.

Analysis of single ingredient tablets of each drug

Ten tablets of Zyrtec^{*} for CTZ or Ventolin^{*} in case of SAL have pulverized accurately and mixed well. An accurately weighed quantity of 20 mg of each drug was transferred to 100 ml volumetric flasks, 80 ml methanol was added and sonicated for 10 min. Solutions were completed with methanol to the volume then filtered. A further dilution was performed to obtain the working concentration range after adding IS. The mean percent content was calculated.

Results and Discussion

The present study describes a simple and reliable HPLC method for the simultaneous determination of CTZ and SAL in their coformulated tablets. After optimization of the chromatographic conditions, CTZ was well separated from SAL with in short retention time (less than 5 min), with a higher number of theoretical plates at pH 3.5 with UV detection at 230 nm (Figures 2 and 3).

Optimization of chromatographic conditions

Typical chromatogram of CTZ and SAL is illustrated in Figure 2. To achieve good separation in short run time, chromatographic conditions were optimized. The studied compounds exhibited

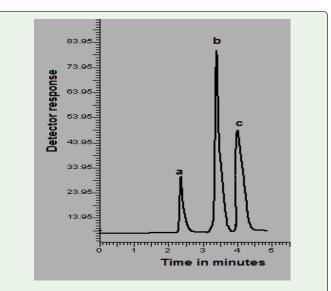


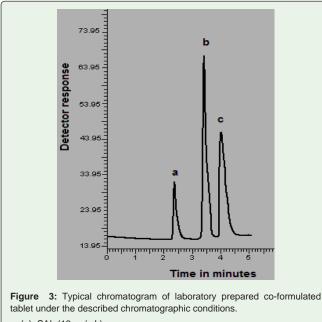
Figure 2: Typical chromatogram of laboratory prepared mixture under the described chromatographic conditions:

(a): SAL (12 μg/mL)
(b): NMS (IS) (20 μg/mL)
(c): CTZ (30 μg/mL)

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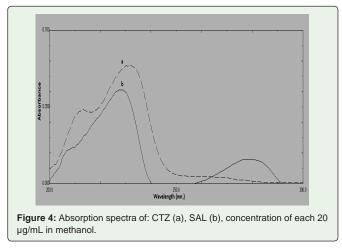
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(a): SAL (12 µg/mL)

(b): NMS (IS) (20 µg/mL)

(C): CTZ (30 µg/mL)



maxima in their spectra at 211 and 231nm for CTZ and 228 nm and 278 nm for SAL. Therefore, the UV detection wavelength was selected to be 230 nm that allowed simultaneous determination of the two drugs with suitable sensitivity (Figure 4).

Different columns were tried to choose the most suitable one for separation of the two drugs, this includes Shimadzu VP-ODS C18 column (250 mm), Shimadzu VP-ODS C18 column (150 mm) and Shim-pack CLC C8 column (250 mm).The first column allowed

Parameter		No. of theoretical plates (N)		Resolution (R_)	Tailing factor (T)		Capacity Factor(K')		Selectivity
		CTZ SAL			CTZ SAL		CTZ	SÁL	factor (α)
	2.5	1156	829	4.3	1	1.25	3.1	0.24	13
	3.5	2600	1225	5.006	1	1.13	6.6	1.5	4.4
pH of the mobile phase	4	1296	1056	4.98	1.3	1.5	1.17	0.22	5.32
	5	1032	987	3.98	1.31	1.51	2.1	0.6	3.4
Conc. of phosphate buffer	0.05	1600	1089	5.423	1.02	1.2	3	0.1	30
	0.1	2600	1225	5.006	1	1.13	6.6	1.5	4.4
	0.2	2381	1225	6.2	1.25	1.04	3.36	0.25	13.4
	50%	792	1024	6.3	2.3	2.49	1.2	0.13	9
Conc. of	70%	1995	924	5.568	1.7	1.25	3.1	0.24	13
methanol (%v/v)	80%	2600	1225	5.006	1	1.13	6.6	1.5	4.4
	90%	797	1344	2.795	1.14	0.99	1.5	0.5	3
	propanol	1251	984	3.007	2.52	1.28	0.67	0.19	3.46
Type of organic modifier 80%, v/v	acetonitrile	1212	1175	1.298	1.54	1.53	0.45	0.31	1.45
0070, 171	methanol	2600	1225	5.006	1	1.13	6.6	1.5	4.4
	0.8	1394	1024	4.79	1.31	1.61	1	0.14	7.05
Flow rate (mL/min)	1	2600	1225	5.006	1	1.13	6.6	1.5	4.4
(m=/mm)	1.2	1277	887	2.309	1.07	1.25	1	0.15	6.6

Table 1: Optimization of chromatographic conditions for HPLC determination of the studied drugs.

Number of theoretical plates (N) = $5.54(\frac{t_R}{W_{h/2}})^2$

Resolution (R_s) = $\frac{2\Delta t_R}{W_1+W_2}$

Tailing factor (T) = $\frac{W_{0.5}}{2f}$

Selectivity factor (Relative retention) (a) = $\frac{t_{R2} - t_m}{t_{R1} - t_m}$ Capacity factor (K') = $\frac{t_R - t_m}{t_{R1}}$ peaks separations with good resolution while the second and the third columns yielded overlapped peaks.

To enhance resolution, efficiency and for good separation, modifications were made in the mobile phase composition including the organic modifier type and ratio, mobile phase pH and ionic strength of the buffer, the results were further explained in Table 1.

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	HPLC	Method
Parameter	СТΖ	SAL
Linearity range (µg/mL)	5.0-50.0	4.0-80.0
Intercept (a)	-0.081	0.192
Slope (<i>b</i>)	0.035	0.01
Correlation coefficient (r)	0.9999	0.9999
S.D. of residuals (S _{y/x})	6.633 x 10 ⁻³	6.66 x 10 ⁻³
S.D. of intercept (S _a)	4.515 x 10 ⁻³	4.866 x 10 ⁻³
S.D. of slope (S _b)	1.707 x 10 ⁻⁴	1.013 x 10 ⁻⁴
S.D.	1.64	0.35
% RSD ª	1.64	0.36
% Error ^b	0.67	0.58
LOD (µg/mL)°	0.57	2.004
LOQ (µg/mL)ª	1.9	4.88

^a Percentage relative standard deviation.

^b Percentage relative error.

° Limit of detection.

Mean

± S.D.

t

F

С

^d Limit of quantitation.

Samuarund		HPLC Method		comparison methods ^{4,11}				
Compound	Amount taken (µg/mL)	Amount found (µg/mL)	% Found	Amount taken	Amount found	% Found		
	5	5.097	101.94	5	4.98	99.58		
	7	7.11	101.63	7	7.04	100.59		
СТХ	10	9.83	98.29	9	8.98	99.77		
012	25	24.89	99.54					
	30	29.67	98.89					
	50	50.03	100.06					
Mean			100.06			99.98		
± S.D.			1.64			0.58		
t			0.087					
F			7.5					
	4	3.9	97.5	10	10.1	101.01		
	10	.9.8	98	20	19.79	98.99		
	35	35.1	100.29	30	830.1	100.34		
SAL	50	49.5	99					
	60	60.9	101.5					

98.9

99.2

0.35

0.95

2.07

79.1

Table 2: Analytical data of CTZ and SAL determination by the proposed method.

To determine the suitable organic modifier, methanol, acetonitrile and n- propanol were used. It was found that acetonitrile and n-propanol showed overlapping peaks of the two drugs. Methanol was chosen for well separated peaks with an increased number of theoretical plates.

The result of altering mobile phase composition was checked using different mobile phases in which methanol ratio varies from 50 to 90% v/v. It was found that ratio 50-70% v/v methanol showed broad CTZ peak. Ratio more than 80% v/v methanol resulted in overlapped peaks. The ratio of (80:20 v/v) methanol: phosphate buffer resulted in well-separated peaks within a reasonable resolution time and with the higher theoretical plates.

The effect of pH of the mobile phase was studied over the range of 2.5-5.5. The peak area ratio of CTZ and SAL decreased with pH higher than 3.5 accompanied by decreasing in the number of theoretical plates. At pH lower than 3.5 there were overlapping between the solvent front and SAL peak. So, pH 3.5 was found to be the most suitable one.

The influence of ionic strength of phosphate buffer was investigated with mobile phases containing 0.05- 0.2 M phosphate buffer. 0.1 M phosphate buffer was selected as it gave well resolved peaks with a higher number of theoretical plates.

Drotroverin. ketoconazole guaifenesin, ambroxol metoclopramide, and nimesulide were tested for the choice of IS.

N.B. Each result is the average of three separate determinations.

The value of tabulated t and Fare2.20and 19.29, respectively at $P = 0.05^{20}$.

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100.11

1.437

Descent		In	tra-day		Inter-day				
Parame	eters	x ±S.D	% RSD	% Error	x ±S.D	% RSD	% Error		
	10.0	98.95 ± 0.51	0.51	0.30	99.68 ± 1.06	1.06	0.61		
SAL (µg/mL)	20.0	99.05 ± 1.42	1.44	0.83	99.57 ± 0.59	0.59	0.34		
	50.0	98.30 ± 0.62	0.64	0.37	99.92 ± 0.39	0.39	0.23		
	25.0	98.20 ± 0.30	0.31	0.18	99.6 ± 1.08	1.09	0.63		
CTZ (µg/mL)	30.0	97.87 ± 0.81	0.83	0.48	100.1 ± 0.90	0.90	0.52		
	50.0	98.57±1.48	1.50	0.86	98.38 ± 0.82	0.84	0.48		

Table 4: Precision data of the proposed method.

N. B. Each result is the average of three separate determinations.

Nimsulide was found to be the most appropriate one yielding wellresolved peak from CTZ and SAL peaks with a suitable retention time (3.5min).

Method Validation

The studied method was validated according to ICH guidelines19 through determination of linearity, range, limits of detection and quantitation, accuracy, precision, robustness, selectivity, system suitability and solutions stability.

A linear relationship was investigated between peak area ratios drug/IS and drug concentration in µg /mL under optimum chromatographic conditions. The concentration ranges were 5.0-50.0 and 4.0-80.0 µg/mL for CTZ and SAL respectively. Statistical analysis of data showed higher values of correlation coefficient (r value > 0.999) and small values of intercept (S_a), slope (S_b), the standard deviation of residuals (S_{y/x}), relative standard deviation and standard error as presented in Table 2. The regression equations for CTZ and SAL are:

y = 0.0349x - 0.0813 for CTZ

y = 0.01x + 0.1921 for SAL

The Limit of Quantitation (LOQ) and the Limit of Detection (LOD) were determined according to ICH recommendations [19]. The results were represented in Table 2.

Table 5: Application of the proposed and comparison methods for determination of the studied drugs in different laboratory prepared mixtures in different pharmaceutical ratios.

Parameter			Proposed method			Comparison methods ^{4,11}				
	Amoun (mg/			t found /mL)	% Found		Amount taken (mg/mL)		% Found	
	CTZ	SAL	CTZ	SAL	CTZ	SAL	CTZ	SAL	CTZ	SAL
HPLC	10	4	9.917	3.947	99.17	98.68	6.25	2.5	101.54	100.56
Method	25	10	25	10.105	100	101.05	7.5	3	100.51	100.63
	30	12	29.667	11.947	98.89	99.56	8.75	3.5	100.22	99.6
	20	20	19.917	20	99.59	100	9	9	100	98.39
	15	30	14.833	29.95	98.89	99.82	5	10	99.24	101.27
Mean					99.31	99.82			100.3	100.09
± S.D.					0.59	1.68			0.73	1.12
t					0.708	1.224				
F					1.82	10.78				

N. B. Each result is the average of three separate determinations.

The value of tabulated *t* and *F* are 2.13 and 6.40, respectively at $P = 0.05^{20}$.

Table 6: Application of the proposed and comparison methods for determination of the studied drugs in their laboratory prepared co-formulated tablets.

		Proposed method						Comparison methods ^{4,11}			
Parameter		it taken /mL)		nt found /mL)	% F	ound		nt taken /mL)	% Fo	und	
	CTZ	SAL	CTZ	SAL	CTZ	SAL	CTZ	SAL	CTZ	SAL	
HPLC	10	4	10.17	3.962	101.7	99.05	6.25	2.5	101.33	98.92	
Method	25	10	24.92	10	99.67	100	7.5	3	97.8	97.77	
	30	12	30.5	11.89	101.7	99.04	8.75	3.5	100.95	99.06	
Mean					101	99.36			100.02	98.58	
± S.D.					1.41	0.55			2.52	0.71	
t					0.75	1.5					
F					2.87	1.65					

N.B. Each result is the average of three separate determinations.

The value of tabulated t and F are 2.78 and 19.00, respectively at $P = 0.05^{20}$.

Table 7: Application of the proposed and comparison methods for determination of the studied drugs in their single ingredient commercial tablets.

Dosage Form	H	IPLC Method		Compariso	n methods 4,11
	Amount taken (μg/mL)	Amount found (μg/mL)	%Found	Amount Found (μg/mL)	% Found
Zyrtic [®] tablets 10mg	6	6.023	100.38	5.892	99.10
CTZ/tablet Mean ± S.D. t F	8	7.955	99.44	7.108	98.65
	10	10.023	100.23	9.98	100.00
Mean			100.02		99.24
± \$.D.			0.51		0.68
t			1.56		
F			1.86		
Ventolin [®] tablets 2 mg	10	10.105	101.05	10.082	100.82
SAL/tablet	20	20.105	100.53	19.837	99.19
	30	30.263	100.88	30.082	100.27
Mean			100.82		100.09
± S.D			0.37		1.15
t			1.45		
F			9.78		

N.B. Each result is the average of three separate determinations.

The value of tabulated t and F are 2.78 and 19.00, respectively at $P = 0.05^{20}$.

The comparison method for CTZ [4] determination in bulk and formulation involved the use of HPLC technique using methanol: 0.01 M disodium hydrogen phosphate buffer (60: 40 v/v), withC18 column and UV detection at 217 nm. An HPLC method [11] was used as a comparison method for SAL determination using acetonitrile: phosphate buffer (65:35 v/v) as a mobile phase. Column C18 was used at 235 nm. The results were compared in terms of accuracy and precision using Student t-test and the variance ratio F-test [20]. Nosignificant difference was observed (Table 3).

Intraday precision and Inter-day precision analyses were determined using the proposed procedures and as shown in Table 4.

The ability of the method to determine CTZ and SAL in their pharmaceutical co-formulated tablet without the interference of additives showed the selectivity of this method (Figure 3).

Applications

Laboratory prepared mixture analysis

Laboratory prepared mixture of CTZ and SAL was analyzed using the proposed method (Figure 2) and the results were compared statistically with those of comparison methods [4,11] as indicated in Table 5.

Dosage form analysis

The suggested procedures were additionally utilized for the determination of CTZ and SAL in their laboratory prepared binary and single ingredient commercial tablets without interference from the excipients. Statistical comparison of the results with that obtained from the comparison methods4,11 using Student t-test and variance ratio F- test20 revealed a non-significant difference in terms of accuracy and precision (Tables 6 and 7).

Conclusion

Our target was to develop new HPLC method for simultaneous determination of CTZ and SAL in their ratio (2.5:1). The suggested method was found to achieve simplicity, accuracy, and time-saving. It was applied successfully for determination of CTZ and SAL in their laboratory prepared co-formulated tablets in short analysis time (less than 5min).

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