



Effect of a multi-carbohydrase and phytase complex on the ileal and total tract digestibility of nutrients in cannulated growing pigs

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Abstract

The current study evaluated the effect of a multi-carbohydrase and phytase complex (MCPC) on ileal and total tract nutrient digestibility in growing pigs. A total of 8 male pigs (initial BW = 30.7±1.1 kg) have been surgically fitted with a T-cannula at the distal ileum and randomly allotted to 4 groups. The experiment has been conducted according to a 4×4 Latin square design, each period lasting 10 days. Pigs were fed 4 experimental diets, which consisted of 2 basal diets (BD1, low phytate; BD2, high phytate) with or without MCPC supplying at least 1800 U xylanase, 6600 U α-arabinofuranosidase, 1244 U β-glucanase, and 1000 U phytase per/kg corn-soybean based diet. High phytate diet reduced ($P < 0.05$) the apparent ileal digestibility (AID) of crude protein by 1.4% and the apparent total tract digestibility (ATTD) of organic matter, crude protein and gross energy by 1.7, 2.3 and 1.9%, respectively, and tended to decrease ($P = 0.10$) the ATTD of Ca by 17.3%, relative to low phytate diet. Dietary supplemental MCPC increased ($P < 0.05$) AID of P and Ca by 34.2% and 31.1% for BD1 and 26.7% and 41.3% for BD2, respectively, as well as increased ($P < 0.05$) ATTD of crude fat, P, and Ca by 1.4%, 45.6% and 9.6% for BD1 and 3.1%, 66.0% and 52.7% for BD2, respectively. MCPC addition did not significantly increase AID and (or) ATTD of crude protein, organic matter and starch. A trend to enhanced protein digestibility was only observed in low phytate BD1 diet. In conclusion, dietary supplemental enzymes rich in MCPC is effective in improving the AID of P and Ca and ATTD of crude fat, P, and Ca.

Keywords: Enzyme, Fecal digestibility, Ileal digestibility, Nutrient, Phytate, Pig

INTRODUCTION

The major storage form of phosphorus (P) in many plant feeds is phytate (myo-inositol hexaphosphate, IP₆) (Lindberg et al., 2007). Since the monogastric livestock lack phytase in their

gastrointestinal tracts, IP₆ can barely be used as P source for animals. In addition, IP₆ is capable of chelating minerals including Ca, Fe, Zn, Mn, and Cu and forming insoluble complexes, and any method enhancing P availability may increase the availability of these elements in feed as well (Pagano et al., 2007). In addition, IP₆ can also bind to protein and reduce the utilization of the proteins and amino acids (Kiarie et al., 2010). Microbial phytase has been widely used as a feed additive for domestic animals to improve utilization of IP₆ from plant feeds (Lei et al., 2013). Most of the IP₆ is in the indigestible fibrous part of cereal grains (Reddy et al., 1982) which also contains non-starch polysaccharides (NSP) that can reduce the nutrient digestion (Passos et al., 2015). Dietary supplementation of NSP degrading carbohydrases might also be beneficial for the availability of P, proteins and amino acid (Lindberg et al., 2007; Passos et al., 2015). However, results for phytase and carbohydrase combinations on nutrient utilization have been inconsistent (Selle et al., 2003; Kim et al., 2005; Woyengo et al., 2008).

Therefore, the aim of the current study was designed to evaluate the effects of the newly developed multi-carbohydrase

Submitted: 12 June 2020 | **Accepted:** 18 July 2020 | **Published:** 22 July 2020

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Citation: Yang JC, Zhang L, Huang YK, Ma R, Wang L, et al. (2020) Effect of a multi-carbohydrase and phytase complex on the ileal and total tract digestibility of nutrients in cannulated growing pigs. *Int J Anim Sci* 4: 6.



and phytase complex (MCPC) supplementation of a corn-soybean based diet on the ileal and total tract digestibility of nutrients in growing pigs fed low and high levels of phytate.

MATERIALS AND METHODS

Pigs, diets, and sample collection

Our animal protocol was approved by the Institutional Animal Care and Use Committee of Huazhong Agricultural University, China. A total of 8 male pigs (Duroc × Large White × Landrace; body weight at 30.7±1.1 kg) have been surgically fitted with a T-cannula at the distal ileum and randomly allotted to 4 groups. Pigs have been housed in individual metabolic crate that allowed freedom of movement and allowed a 14-d recovery period. After the recovery, pigs were fed one of the 4 experimental diets, which consisted of 2 basal diets (BD1, low phytate; BD2, high phytate; Table 1), with or without MCPC (Rovabio Advance Phy, Adisseo France S.A.S., France) supplying at least 1800 U xylanase, 6600 α-arabinofuranosidase, 1244 U β-glucanase, and 1000 U phytase per/kg diet. All diets contain 0.5% titanium dioxide (TiO₂) as an indigestible marker and were fed as mash. The experiment has been conducted according to a double 4×4 Latin square design. The pigs have been fed with a daily feed allowance at 4% body weight in 2 equal meals at 0800 and 1600 and adjusted every week. Pigs were allowed free access to water. Each period lasted 10 days, including 5 days of adaptation, followed by 3 days of feces collection and 2 days of ileal digesta collection. Feces have been collected using plastic bags attached to the skin around the anus. Digesta samples have been collected for 2 d using bags

containing diluted formic acid attached to the opened cannula barrel from 08:00 to 20:00 h (Yáñez et al., 2011). Collected feces and digesta samples have been pooled for each pig within experimental period and frozen at -20°C.

Chemical analyses

The organic matter, gross energy, crude protein, crude fat, starch, P, Ca, and ash in the BD were analyzed according to previous studies (Noblet et al., 1994; Varley et al., 2011; Conde-Aguilera et al., 2016). Briefly, the gross energy of samples was determined using an adiabatic oxygen bomb calorimeter according to procedures outlined by the Association of Official Analytical Chemists (AOAC, 1980), and starch content was measured using the Ewers polarimetric method. In the AOAC (1990) frame, the content of Ca and P is determined by spectrophotometry; the content of crude protein is measured by Kjeldahl method; the content of crude fat is determined by diethyl ether extraction; and determined the content of ash and organic matter after completely burned. The AID and ATTD of components in the diet were calculated using the equation as described before (Yáñez et al., 2010).

Statistical analysis

Data has been subjected to ANOVA using the General Linear Model using SAS (SAS Inst. Inc., Cary, NC). The statistical model included the period, animal number, phytate level, MCPC and phytate × MCPC interaction. Tukey test was used in order to compare the means. Differences were considered significant at $P < 0.05$.

Table 1: Composition of basal diets and nutritional contents¹.

Ingredients	Percentage (%)		Calculated nutrients	Content	
	BD1	BD2		BD1	BD2
Corn	58.00	50.62	NE, MJ/kg	9.80	9.80
Corn distillers	15.00	15.00	Crude protein	17.62	15.80
Rice bran	2.53	15.00	Crude fat	5.30	7.12
Soybean meal	13.25	6.00	Dig. Lysine	0.95	0.95
Rapeseed meal	7.00	7.00	Dig. Methionine	0.28	0.31
Soybean oil	1.09	2.94	Dig. Methionine+cysteine	0.52	0.52
DL-methionine	0.00	0.05	Ca	0.66	0.66
L-lysine	0.42	0.59	Digestible P	0.31	0.31
L-threonine	0.03	0.11	Phytic P ²	0.31	0.46
Tryptophan	0.02	0.04	Starch	39.03	38.06
Calcium carbonate	1.09	1.10	Soluble arabinoxylan ²	0.77	0.66
CaPO ₃	0.20	0.19	Insoluble arabinoxylan ²	5.60	5.01
Salt	0.37	0.37	Total arabinoxylan ²	6.37	5.67
Titanium oxide	0.50	0.50			
Premix ²	0.50	0.50			

¹BD = based diet; BD1= low phytate; BD2 = high phytate.

²Premix provided/kg diet: retinyl acetate, 10000 IU; cholecalciferol 2500 IU; dl-α-tocopheryl acetate, 50 IU; menadione, 5.0 mg; thiamin, 2.0 mg; riboflavin, 5.0 mg; pantothenic acid, 12.0 mg; pyridoxine, 10.0 mg; niacin, 30.0 mg; biotin, 0.2 mg; folic acid, 1.5 mg; cyanocobalamin 0.05 mg; choline chloride 1500 mg; iron, 100 mg; copper, 20 mg; manganese, 25 mg; zinc, 100 mg; selenium, 0.3 mg; iodine, 0.3 mg.

²Calculated values.



Table 2: The analyzed nutrient composition of the based diets¹.

Nutrients	BD1	BD2
Gross energy, MJ/kg	16.70	17.40
Crude protein, %	18.48	16.85
Crude fat, %	5.44	7.73
Ash, %	5.14	5.23
Starch, %	44.75	45.61
P, %	0.52	0.61
Ca, %	0.70	0.68

¹BD = based diet; BD1= low phytate; BD2 = high phytate.

RESULTS

The analyzed nutrient composition of BD is presented in Table 2, including gross energy, crude protein, crude fat, starch, P, and Ca and are in agreement with their calculated values in Table 1. The AID of crude protein was significantly affected by dietary phytate content (Table 3). High phytate diet decreased ($P < 0.05$) the AID of crude protein by 1.4% units relative to low phytate diet in -MCPC groups. Dietary supplemental enzymes significantly affected AID of P, and Ca (Table 3). Specifically, dietary added MCPC increased the AID of P and Ca by 34.2% and 26.7% and 31.1% and 41.3% at low and high phytate level, respectively. However, no significant interaction between phytate level and MCPC has been observed on AID of these nutrients.

The ATTD of organic matter, crude protein, crude fat, Ca and gross energy were significantly affected by dietary phytate content (Table 3). High phytate diet decreased ($P < 0.05$) organic matter, crude protein, gross energy and Ca by 1.7, 2.3, 1.9 and 17.3%, respectively, while it increased ($P < 0.05$) crude fat by 2.8% relative to the low phytate diet in the -MCPC groups.

Dietary supplemental enzymes significantly affected (Table 3) ATTD of crude fat, P, and Ca. Specifically, dietary added enzymes increased ($P < 0.05$) the ATTD of crude fat, P, and Ca by 1.4 and 3.1%, 45.6% and 66.0%, 9.6 and 52.7% at low and high phytate levels, respectively. However, no significant interaction between phytate level and MCPC has been observed on these nutrients and no treatment effect has been observed on starch ATTD.

DISCUSSION

Current study showed that the ATTD of organic matter, crude protein, gross energy, and Ca and the AID of crude protein were significantly reduced by dietary phytate content in growing pigs, as observed by Woyengo et al. (2012) and Kahindi et al. (2015). Phytate may bind to glucose, starch and amino acids and may decrease the amylase and protease enzyme activities in the small intestine reducing thereby the digestibility and utilization of nutrients (Selle et al., 2000; Pagano et al., 2007; Kiarie et al., 2010; Selle et al., 2012; Woyengo and Nyachoti, 2013). It is generally accepted that by breaking down phytate, phytase limit the insoluble complex formation between phytate and nutrients, thus improving nutrient digestibility (Lei et al., 2013; Cowieson et al., 2017; Zouaoui et al., 2018). Interestingly, the current study showed that dietary supplemental enzymes rich in phytase and multi-carbohydrase increased the AID of P and Ca, as well as the ATTD of crude fat, P, and Ca in both low and high phytate diets. This could be explained by the enzyme degradation of IP₆ and polysaccharides in the cell walls in cereals and storage cell contents in protein meals and reduction of digesta viscosity (Lei et al., 2013; Passos et al., 2015). These outcomes were similar with previous studies, which have shown that dietary supplemental either phytase, NSP degrading enzymes, or both improved the AID and ATTD of crude fat, P, and/or Ca in swine (Radcliffe et al., 2006; Woyengo et al., 2008; Selle et al., 2009; Guggenbuhl et al., 2012; Torres-Pitarch et al., 2017). Moreover, the present

Table 3: Effect of phytate level and Enzymes on the apparent total tract digestibility of nutrients.

	Low phytate		High phytate		<i>P</i> -value ¹			<i>R</i> ²
	-MCPC	+MCPC	-MCPC	+MCPC	Phytate	MCPC	Phytate x MCPC	
AID, % ²								
Organic matter	82.59	82.62	82.77	83.38	0.14	0.31	0.35	0.73
Crude protein	82.35	83.69	81.23	81.87	0.04	0.14	0.60	0.62
P	47.24	63.40	50.60	64.09	0.17	< 0.01	0.36	0.88
Ca	47.60	62.40	43.67	61.69	0.33	< 0.01	0.51	0.85
ATTD, % ²								
Organic matter	85.82	85.49	84.33	84.57	< 0.01	0.89	0.42	0.84
Starch	91.85	91.56	91.75	91.97	0.72	0.93	0.55	0.81
Crude protein	81.52	83.28	79.63	79.92	0.001	0.15	0.30	0.82
Crude fat	79.11	80.22	81.29	83.82	< 0.001	< 0.01	0.22	0.91
Gross energy	83.19	82.56	81.64	82.05	0.03	0.81	0.24	0.84
P	32.67	47.57	27.55	45.72	0.20	< 0.01	0.54	0.78
Ca	49.59	54.34	40.99	62.60	0.10	< 0.01	0.28	0.95

¹Data (n=32) were subject to variance analysis with fixed effect phytate (n=2), MCPC (n=2) and interaction (n=4)

²AID = apparent ileal digestibility; ATTD = apparent total tract digestibility; MCPC = multi-carbohydrase and phytase complex; -, diet without added MCPC; +, diet with added MCPC.



study showed that dietary supplementation of phytase plus multi-carbohydrase rich in xylanase, α -arabinofuranosidase and β -glucanase resulted in higher increased ATTD of P (66.0%) and Ca (52.7%) in higher phytate diet than dietary supplementation only of phytase at 1000 FTU/kg which resulted in increased ATTD of P (32.2%) and Ca (15.5%) (She et al., 2018). These outcomes are consistent with a previous study that showed dietary supplementation of phytase and xylanase together displayed better results of AID and ATTD of P and Ca than dietary supplementation of xylanase alone (Lindberg et al., 2007). These findings could be explained by a complementary effect between multi-carbohydrase and phytase on the digestibility of both minerals. Nevertheless, several inconsistent scenarios were observed in the present study. Dietary supplemental enzymes did not affect the AID and ATTD of crude protein as reported by Lindberg et al. (2007) and Woyengo et al. (2008). Also, the ATTD of starch was not affected neither by phytate level nor by MCPC supplementation. However, some other studies showed that dietary supplementation of either phytase or NSP-degrading carbohydrase alone or in combination can improve the digestibility of nutrients (Zeng et al., 2018; Lee et al., 2019; Sun et al., 2020). This discrepancy could also be due to different experimental conditions, including diet compositions, doses and types of enzymes, and ages of pigs. Moreover, no interaction was observed between phytic P level and MCPC on the digestibility of nutrients in this study. However, the protein digestibility tended to be increased with MCPC on low phytate diets (+1.6 and +2.2 % on AID and ATTD of protein respectively) but surprisingly not on high phytate diet. This observation might also be due to the large difference between low and high phytate diets which were not based only on phytate level difference but also on ingredients used such as high rice bran content as well as on crude fat and crude protein contents. This result may also indicate that MCPC is less impacted by level of phytic P ranging from 0.31 to 0.46%, probably due to its high active phytase at pH 3.0 present within the MCPC (Menezes-Blackburn et al., 2015), which lead to more efficient breakdown of phytate in the stomach and reduced the formation of insoluble complexes between phytate and nutrients.

CONCLUSION

In summary, the current study found that the dietary supplement of an enzymatic cocktail with phytase and multi-carbohydrase improved the ileal and total tract digestibility of P, Ca, crude fat in growing pigs and could be used as a promising enzymes product to reduce the negative effects of phytate in practice.

CONFLICT OF INTEREST

All authors declare that there are no conflicts of interests.

ACKNOWLEDGEMENTS

This work was supported in part by the National Key Research and Development Program of China, Project (2016YFD0501207 and 2018YFD0500601); Innovation Group of Hubei Natural Science Foundation (2018CFA020) and a research gift from Adisseo France S.A.S.

ANIMAL WELFARE STATEMENT

The experimental design and procedures in this study were reviewed and approved by department of Animal Nutrition and Feed Science, College of Animal Science and Technology, Huazhong Agricultural University, Wuhan, Hubei, China.

All animal experimental procedures were performed according to the Institutional Animal Care and Use Committee of Huazhong Agricultural University.

References

1. AOAC. (1980). Official Methods of Analysis. 13th Edition, Association of Official Analytical Chemists, Gaithersburgs, MD. <https://doi.org/10.1021/ac50052a726>
2. Bedford MR & Schulze H. Exogenous enzymes for pigs and poultry (1998). *Nutr Res Rev*, 11(1), 91–114. <https://doi.org/10.1079/NRR19980007>
3. Casas, G.A., and Stein, H.H. (2016). Effects of microbial xylanase on digestibility of dry matter, organic matter, neutral detergent fiber, and energy and the concentrations of digestible and metabolizable energy in rice coproducts fed to weanling pigs. *J ANIM SCI*, 94, 1933-1939. <https://doi.org/10.2527/jas.2015-0064>
4. Cozannet P, Kidd MT, Montanhini Neto R, Geraert P-A. (2017) Next-generation non-starch polysaccharide-degrading, multi-carbohydrase complex rich in xylanase and arabinofuranosidase to enhance broiler feed digestibility. *Poult Sci*, 96(8), 2743–2750. <https://doi.org/10.3382/ps/pex084>
5. Greiner R, Haller E, Konietzny U, Jany KD. (2010). Purification and Characterization of a Phytase from *Klebsiella terrigena*. *Iubmb Life*, 40(1), 83–91. <https://doi.org/10.1006/abbi.1997.9942>
6. Guggenbuhl P, Waché Y, Simoes Nunes C, Fru F. (2012). Effects of a 6-phytase on the apparent ileal digestibility of minerals and amino acids in ileorectal anastomosed pigs fed on a corn-soybean meal-barley diet. *J Anim Sci*, 90 Suppl4(13), 182–184. <https://doi.org/10.2527/jas.53892>
7. Hill BE, Sutton AL, Richert BT. (2009). Effects of low-phytic acid corn, low-phytic acid soybean meal, and phytase on nutrient digestibility and excretion in growing pigs. *J Anim Sci*, 87(4), 1518–1527. <https://doi.org/10.2527/jas.2008-1219>
8. Humer E, Schwarz C, Schedle K. (2015). Phytate in pig and poultry nutrition. *J Anim Physiol Anim Nutr*. 99(4), 605–25. <https://doi.org/10.1111/jpn.12258>
9. Kahindi RK, Thacker PA, Nyachoti CM. (2015) Nutrient digestibility in diets containing low-phytate barley, low-phytate field pea and normal-phytate field pea, and the effects of microbial phytase on energy and nutrient digestibility in the low and normal-phytate field pea fed to pigs. *Anim Feed Sci Technol*, 203,79–87. <https://doi.org/10.1016/j.anifeedsci.2015.02.009>
10. Kiarie E, Owusu-Asiedu A, Simmins PH, Nyachoti CM. (2010). Influence of phytase and carbohydrase enzymes on apparent ileal nutrient and standardized ileal amino acid digestibility in growing pigs fed wheat and barley-based diets. *Livest Sci*, 134(1–3), 85–87. <https://doi.org/10.1016/j.livsci.2010.06.105>
11. Kim JC, Simmins PH, Mullan BP, Pluske JR. (2005). The effect of wheat phosphorus content and supplemental enzymes on digestibility and growth performance of weaner pigs. *Anim Feed Sci Technol*, 118(1–2):139–152. <https://doi.org/10.1016/j.anifeedsci.2004.08.016>



12. Kim JC, Sands JS, Mullan BP, Pluske JR. (2008). Performance and total-tract digestibility responses to exogenous xylanase and phytase in diets for growing pigs. *Anim Feed Sci Technol*, 142(1–2), 163–172. <https://doi.org/10.1016/j.anifeedsci.2007.07.004>
13. Lee JW, Patterson R, Rogiewicz A, Woyengo TA. (2019). Nutrient digestibility of multi-enzyme supplemented low-energy and AA diets for grower pigs. *J Anim Sci*, 97(7), 2979–2988. <https://doi.org/10.1093/jas/skz178>
14. Lei XG, Weaver JD, Mullaney E, Ullah AH, Azain MJ. (2013). Phytase, a new life for an “old” enzyme. *Annu Rev Anim Biosci*, 1(1), 283–309. <https://doi.org/10.1146/annurev-animal-031412-103717>
15. Liao SF, Sauer WC, Kies AK, Zhang YC, Cervantes M, He JM. (2005). Effect of phytase supplementation to diets for weaning pigs on the digestibilities of crude protein, amino acids, and energy. *J Anim Sci*, 83(3), 625–633. <https://doi.org/10.2527/2005.833625x>
16. Lindberg JE, Lyberg K, Sands J. (2007). Influence of phytase and xylanase supplementation of a wheat-based diet on ileal and total tract digestibility in growing pigs. *Livest Sc*, 109(1–3), 268–270. <https://doi.org/10.1016/j.livsci.2007.01.114>
17. Menezes-Blackburn D, Gabler S, Greiner R. (2015). Performance of Seven Commercial Phytases in an in Vitro Simulation of Poultry Digestive Tract. *J Agric Food Chem*, 63(27), 6142–6149. <https://doi.org/10.1021/acs.jafc.5b01996>
18. Noblet, J., and Shi, X.S. (1994). Effect of body weight on digestive utilization of energy and nutrients of ingredients and diets in pigs. *Livestock Production Science*, 37, 323–333. [https://doi.org/10.1016/0301-6226\(94\)90126-0](https://doi.org/10.1016/0301-6226(94)90126-0)
19. Nortey TN, Patience JF, Sands JS, Trottier NL, Zijlstra RT. (2008). Effects of xylanase supplementation on the apparent digestibility and digestible content of energy, amino acids, phosphorus, and calcium in wheat and wheat by-products from dry milling fed to grower pigs. *J Anim Sc*, 86(12), 3450–3464. <https://doi.org/10.2527/jas.2007-0472>
20. O’Dell BL, Sunde RA. (1997). Handbook of nutritionally essential mineral elements. *CRC Press*. <https://doi.org/10.1201/9781482273106>
21. Pagano AR, Yasuda K, Roneker KR, Crenshaw TD, Lei XG. (2007). Supplemental *Escherichia coli* phytase and strontium enhance bone strength of young pigs fed a phosphorus-adequate diet. *J Nut*, 137(7), 1795–1801. <https://doi.org/10.1093/jn/137.7.1795>
22. Passos AA, Park I, Ferket P, von Heimendahl E, Kim SW. (2015) Effect of dietary supplementation of xylanase on apparent ileal digestibility of nutrients, viscosity of digesta, and intestinal morphology of growing pigs fed corn and soybean meal based diet. *Anim Nutr*, 1(1), 19–23. <https://doi.org/10.1016/j.aninu.2015.02.006>
23. Radcliffe JS, Pleasant RS, Kornegay ET. (2006). Estimating equivalency values of microbial phytase for amino acids in growing and finishing pigs fitted with steered ileo-cecal valve cannulas. *J Anim Sci*, 84(5), 1119–1129. <https://doi.org/10.2527/2006.8451119x>
24. Reddy NR, Sathe SK, Salunkhe DK. (1982). Phytates in legumes and cereals. *Adv. Food Res*, 28, 1–92. [https://doi.org/10.1016/s0065-2628\(08\)60110-x](https://doi.org/10.1016/s0065-2628(08)60110-x)
25. Saleh AA, Kirrella AA, Abdo SE, Mousa MM, Badwi NA, Ebeid TA, et al. (2019). Effects of Dietary Xylanase and Arabinofuranosidase Combination on the Growth Performance, Lipid Peroxidation, Blood Constituents, and Immune Response of Broilers Fed Low-Energy Diets. *Animals*, 9(7), 467. <https://doi.org/10.3390/ani9070467>
26. Selle PH, Cowieson AJ, Cowieson NP, Ravindran V. (2012). Protein-phytate interactions in pig and poultry nutrition: a reappraisal. *Nutr Res Rev*, 25(1):1–17. <https://doi.org/10.1017/s0954422411000151>
27. Selle PH, Ravindran V, Partridge GG. (2009). Beneficial effects of xylanase and/or phytase inclusions on ileal amino acid digestibility, energy utilisation, mineral retention and growth performance in wheat-based broiler diets. *Anim Feed Sci Technol*, (3–4), 303–313. <https://doi.org/10.1016/j.anifeedsci.2009.06.011>
28. Selle PH, Ravindran V, Ravindran G, Pittolo PH, Bryden WL. (2003). Influence of phytase and xylanase supplementation on growth performance and nutrient utilisation of broilers offered wheat-based diets. *Asian-Australasian J Anim Sci*, 16(3), 394–402. <https://doi.org/10.5713/ajas.2003.394>
29. She Y, Sparks JC, Stein HH. (2018). Effects of increasing concentrations of an *Escherichia coli* phytase on the apparent ileal digestibility of amino acids and the apparent total tract digestibility of energy and nutrients in corn-soybean meal diets fed to growing pigs. *J Anim Sci*, 96(7), 2804–2816. <https://doi.org/10.1093/jas/sky152> <https://doi.org/10.1093/jas/sky152>
30. Sun H, Cozannet P, Ma R, Zhang L, Huang Y-K, Preynat A, et al. (2020). Effect of concentration of arabinoxylans and a carbohydrase mixture on energy, amino acids and nutrients total tract and ileal digestibility in wheat and wheat by-product-based diet for pigs. *Anim Feed Sci Technol*, 262, 114380. <https://doi.org/10.1016/j.anifeedsci.2019.114380>
31. Sun LH, Qin T, Liu Y, Zhao H, Xia X, Lei X. (2018). Cloning, expression, and characterization of a porcine pancreatic α -amylase in *Pichia pastoris*. *Anim Nutr*, 4(2), 234–240. <https://doi.org/10.1016/j.aninu.2017.11.004>
32. Torres-Pitarch A, Hermans D, Manzanilla EG, Bindelle J, Everaert N, Beckers Y, et al. (2017). Effect of feed enzymes on digestibility and growth in weaned pigs: a systematic review and meta-analysis. *Anim Feed Sci Technol*, 233, 145–159. <https://doi.org/10.1016/j.anifeedsci.2017.04.024>
33. Varley, P.F., Callan, J.J., and O Doherty, J.V. (2011). Effect of dietary phosphorus and calcium level and phytase addition on performance, bone parameters, apparent nutrient digestibility, mineral and nitrogen utilization of weaner pigs and the subsequent effect on finisher pig bone parameters. *ANIM FEED SCI TECH*, 165, 201–209. <https://doi.org/10.1016/j.anifeedsci.2011.02.017>
34. Viveros A, Centeno C, Brenes A, Canales R, Lozano A. (2000). Phytase and acid phosphatase activities in plant feedstuffs. *J Agric Food Chem*, 48(9), 4009–4013. <https://doi.org/10.1021/jf991126m>
35. Woyengo TA, Akinremi OO, Rosnagel BG, Nyachoti CM. (2012). Performance and total tract nutrient digestibility of growing pigs fed hullless low phytate barley. *Can J Anim Sci*, 92(4), 505–511. <https://doi.org/10.4141/cjas2012-038>
36. Woyengo TA, Ige D V, Akinremi OO, Nyachoti CM. (2016). Performance and nutrient digestibility in growing pigs fed wheat dried distillers’ grain with solubles-containing diets supplemented with phytase and multi-carbohydrase. *Anim Sci J*, 87(4), 570–577. <https://doi.org/10.1111/asj.12461>
37. Woyengo TA, Nyachoti CM. (2013). Anti-nutritional effects of phytic acid in diets for pigs and poultry—current knowledge and directions for future research. *Can J Anim Sci*, 93(1), 9–21. <https://doi.org/10.4141/cjas2012-017>
38. Woyengo TA, Sands JS, Guenter W, Nyachoti CM. (2008). Nutrient digestibility and performance responses of growing pigs fed phytase- and xylanase-supplemented wheat-based diets. *J Anim Sci*, 86(4), 848–857. <https://doi.org/10.2527/jas.2007-0018>
39. Yáñez JL, Beltranena E, Cervantes M, Zijlstra RT. (2011). Effect of phytase and xylanase supplementation or particle size on nutrient



- digestibility of diets containing distillers dried grains with solubles cofermented from wheat and corn in ileal-cannulated grower pigs. *J Anim Sci*, 89(1), 113–123. <https://doi.org/10.2527/jas.2010-3127>
40. Yi JQ, Piao XS, Li ZC, Zhang HY, Chen Y, Li QY, et al. (2013). The effects of enzyme complex on performance, intestinal health and nutrient digestibility of weaned pigs. *Asian-Australasian J Anim Sci*, 26(8), 1181-8. <https://doi.org/10.17221/7339-cjas>
41. Zeng ZK, Li QY, Tian QY, Xu YT, Piao XS. (2018). The combination of carbohydrases and phytase to improve nutritional value and non-starch polysaccharides degradation for growing pigs fed diets with or without wheat bran. *Anim Feed Sci Technol*, 235, 138–46. <https://doi.org/10.1016/j.anifeedsci.2017.11.009>