

Properties and Applications of Plants of
Origanum Sp. GenusJosé María García-Beltrán¹ and María Ángeles Esteban^{1*}¹Department of Cell Biology and Histology, University of Murcia, Spain

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Abstract

The genus *Origanum* consists of different aromatic and medicinal plants some of which are used in folk medicine and as food additives since ancient times. These plants have numerous and varied beneficial properties, among which are antibacterial, antifungal, antioxidant, anti-inflammatory, antitumor and antiviral. While a mixture of components present naturally in these plants confers myriad of benefits phenolic compounds in particular have great importance in biocidal and antioxidants properties. In this review we focus on the genus *Origanum*, discussing the beneficial and probed properties that have potential implications in health-care and dietetics.

Introduction

Phytotherapy is the science that studies the use of plant products for therapeutic purposes [1]. Medicinal plants and phytochemical products have been used for the treatment of various diseases in traditional medicine worldwide not least because of its remedial properties [2,3] and, furthermore, they have been used as human food additives due to their beneficial properties [4] such as biocidal and antioxidants. It is also important to note that currently, about 80% of the world population uses herbs as traditional remedies [2]. However, although these products are natural, they can have side effects, although less than synthetic drugs [1,2].

Among the aromatic and medicinal plants, *Origanum sp.* is an important genus belonging to the *Lamiaceae* family [5]. It consists of forty nine taxa divided into 10 sections [6,7]. Most of its species are distributed around the Mediterranean area [7], Eurasia and the North of Africa [5]. This genus is very appreciated for its volatile oil and it is characterized by a great morphological and chemical diversity [3,6]. Since antique *Origanum sp.* plants have been used in folk medicine and ethnomedicine [5,8] to treat numerous infectious diseases and pains. It is therefore important to ensure that their use is in appropriate dosages because while they confer beneficial properties in therapeutic doses over a short period of time, they however can be toxic if taken in excess [9]. In addition, it has also been demonstrated that *Origanum* species have many properties considered important from a culinary and agricultural point of view. Among them are their ovicidal, herbicidal and insecticidal activities [10,11]. They are also used as food spices to flavor food [12,13]. Furthermore, and thanks to its antimicrobial activity, plant belonging to this genus retards the growth of microorganisms in food and therefore can also be used as food preservative and flavoring agents as well as disinfectants in perfumes and soaps [3]. These plants have all the properties and beneficial activities enumerated due to the presence of different essential oils which are products of the secondary metabolism of plants [3,10]. At present, it has been demonstrated that the oils present in the whole plant give their properties [3,11,13,14]. Although essential oils can be obtained from flowers, buds, seeds, leaves, branches, barks, fruits and roots [15], they are however commonly obtained from the aerial parts of the plants.

In this review we focus on the study of the genus *Origanum*, especially in its beneficial and demonstrated properties. We have wanted to give a detailed account of many beneficial properties of *Origanum* that have potential implications in health-care and dietetics.

Properties of the *Origanum* Genus

As stated earlier, the properties of genus *Origanum* plants are many and varied, thanks to which, man has benefited from their use since ancient times. In this section we present some of the activities attributed to these plants.

Origanum species have many activities related to medical, culinary and agricultural importance. Among the properties related to medical importance are the following: gastrointestinal diseases (such as diarrhea, stomach pain, colic and gastric ulcers), respiratory diseases (e.g. asthma, cough and chest pain) [3,11,16], abnormal menstrual cycles [5], kidney and liver diseases, metabolic, hormonal and neuronal disorders and skin and urogenital system diseases [11]. They have also been used as a sedative and their biocidal properties such as antiparasitic and anthelmintic [3,5], antibacterial, antifungal and antiviral ones [3,17] have been well documented. *Origanum sp.* have also antimutagenic,

antitumor and cytotoxic activities [17-20] as well as antioxidant, anti-inflammatory [14,16,21], antispasmodic, expectorant, carminative and antitussive properties [20]. Furthermore, *Origanum* has been used to treat nausea and rheumatism [16], arthritis, hemorrhoids, sexual diseases, animal bites and poisoning [22] and to control diabetes and obesity [11,23]. These species have been also used as carminatives, diaphoretics and tonics, and as a source of antimicrobial compounds [5].

Due to the fact that there are numerous studies about these plants, only some of them, considered as more representatives, will be discussed in more detail in this review. Furthermore, we will focus only on antibacterial, antifungal, antioxidant, anti-inflammatory, antitumor, antiparasitic, antiviral, antihyperglycemic and anticholinesterase activities.

Antibacterial activity

These plants have antibacterial activities which give them a very important role not only for treating infectious bacterial diseases, but also for using as food preservatives role, as they much delay microbial growth and better preserved food. All the studied species of the genus *Origanum* present antibacterial activity. Curiously, this activity is broad not confined to Gram negative or Gram positive bacteria thus being particularly attractive in the field of medicine. Furthermore, this plant has also activity against plants or trees pathogenic bacteria, and therefore demonstrating relevance and importance in agriculture. The antibacterial activities verified in some species of the genus *Origanum* are shown in Table 1 for Gram positive and Table 2 for Gram negative bacteria.

Infusions of both essential oils and plant extracts (either aqueous or ethanolic) can be used in antimicrobial formulations as they have activity against strains of Gram positive and negative bacteria [9,24,40]. The hydrophobicity of the essential oils allows them to be embedded in the cell and in the mitochondrial membranes and breaks them, making more permeable to throw out contents of the cell [4,15]. Different conditions such as low pH, temperature or oxygen levels enhanced the antibacterial action of essential oils. The antibacterial activity of the essential oils possibly is due to the presence of phenolic compounds such as carvacrol and thymol [10,11,20,25,30,34], and also γ -terpinene [10], linalool, p-cymene, 4-terpineol and β -caryophyllene [15,20,25]. Possibly, this activity is due to the synergism between some of the compounds present in oils [15].

Antifungal activity

As it was previously described for the antibacterial properties, the antifungal properties of these plants are very important, also regarding the possibility of treating infectious diseases and preserving food. Similarly, these plants also affect plants and trees pathogenic fungi that cause heavy losses to farmers, so they are also important for the agricultural sector. The antifungal properties studied in some species of the genus *Origanum* are shown in Table 3.

Origanum sp. essential oils and extracts (both aqueous and ethanolic) of essential oils and/or aerial parts of plants exhibit antifungal activity [45]. This antifungal activity is also due to the presence of thymol and carvacrol [34,49]. Furthermore, the quantity of carvacrol content is correlated to the antifungal activity [3].

Antioxidant activity

The oxidation reactions are very usual in the cells although they can produce Reactive Oxygen Species (ROS) and free radicals that start a series of reactions that finally damage and/or kill the cells. The antioxidants protect the cells and also their components, and thus protect the organism. The antioxidant properties of these plants have also been shown in a large number of studies and those considered as most representatives are shown in Table 4.

Table 1: *Origanum* species with activity against Gram positive bacteria.

Origanum species	Bacteria	References
<i>O. syriacum</i>	<i>Staphylococcus aureus</i>	[3]
	<i>Enterococcus faecalis</i>	[3,10]
	<i>Bacillus brevis</i>	[10]
	<i>Bacillus megaterium</i>	[10]
	<i>Bacillus subtilis</i>	[10]
	<i>Micrococcus luteus</i>	[10]
	<i>Mycobacterium smegmatis</i>	[10]
<i>O. vulgare</i>	<i>Listeria innocua</i>	[19]
	<i>Staphylococcus epidermidis</i>	[9]
	<i>Staphylococcus aureus</i>	[9,24-27]
	<i>Staphylococcus saprophyticus</i>	[24]
	<i>Micrococcus sp</i>	[24]
	<i>Bacillus sp</i>	[25]
	<i>Bacillus cereus</i>	[25]
	<i>Bacillus subtilis</i>	[25]
	<i>Bacillus pumilis</i>	[27]
	<i>Enterococcus faecalis</i>	[28]
	<i>Listeria innocua</i>	[28,29]
	<i>Listeria monocytogenes</i>	[30]
	<i>Bochothrix thermosphacta</i>	[30]
<i>O. dictamnus</i>	<i>Micrococcus luteus</i>	
	<i>Staphylococcus aureus</i>	[13,20,31-33]
	<i>Staphylococcus epidermidis</i>	[13,20,31-33]
	<i>Listeria monocytogenes</i>	[13,33]
	<i>Bacillus cereus</i>	[20]
	<i>Bacillus subtilis</i>	[20]
	<i>Streptococcus faecalis</i>	[20]
	<i>Streptococcus mutans</i>	[31,32]
	<i>Streptococcus viridans</i>	[33]
<i>O. microphyllum</i>	<i>Staphylococcus hominis</i>	[33]
	<i>Bacillus cereus</i>	[20]
	<i>Bacillus subtilis</i>	[20]
	<i>Staphylococcus aureus</i>	[20]
<i>O. libanoticum</i>	<i>Staphylococcus epidermidis</i>	[20]
	<i>Bacillus cereus</i>	[20]
	<i>Bacillus subtilis</i>	[20]
<i>O. majorana</i>	<i>Staphylococcus epidermidis</i>	[20]
	<i>Bacillus cereus</i>	[25]
	<i>Bacillus subtilis</i>	[25]
	<i>Bacillus pumilis</i>	[25]
<i>O. acutidens</i>	<i>Staphylococcus aureus</i>	[25,26]
	<i>Bacillus macerans</i>	[34]
	<i>Bacillus megaterium</i>	[34]
	<i>Bacillus subtilis</i>	[34]
	<i>Clavibacter michiganense</i>	[34]
	<i>Enterococcus faecalis</i>	[34]
<i>O. glandulosum</i>	<i>Staphylococcus aureus</i>	[34]
	<i>Staphylococcus epidermis</i>	[34]
<i>O. minutiflorum</i>	<i>Enterococcus hirae</i>	[35]
	<i>Staphylococcus aureus</i>	[35]
<i>O. minutiflorum</i>	<i>Listeria monocytogenes</i>	[36]
	<i>Staphylococcus aureus</i>	[36]

Table 2: *Origanum* species active against Gram negative bacteria.

Origanum species	Bacteria	Reference
O. syriacum	<i>Pseudomonas aeruginosa</i>	[3,10]
	<i>Escherichia coli</i>	[3,10,37]
	<i>Klebsiella oxytoca</i>	[10]
	<i>Yersinia enterocolitica</i>	[10]
	<i>Klebsiella pneumoniae</i>	[10,38]
	<i>Escherichia coli</i> O157:H7	[38]
	<i>Proteus spp</i>	[38]
	<i>Yersinia enterocolitica</i> O9	[38]
O. vulgare	<i>Brucella melitensis</i>	[39]
	<i>Enterobacter aerogenes</i>	[9]
	<i>Enterobacter sakazakii</i>	[9]
	<i>Proteus vulgaris</i>	[9]
	<i>Escherichia coli</i>	[9,25,27,40]
	<i>Pseudomonas aeruginosa</i>	[9,25,40]
	<i>Salmonella poona</i>	[25]
	<i>Salmonella Enteritidis</i>	[27]
	<i>Salmonella Typhimurium</i>	[27]
	<i>Pseudomonas fragi</i>	[30]
	<i>Salmonella sp.</i>	[30]
	<i>Aeromonas hydrophila</i>	[30]
	<i>Citrobacter spp.</i>	[40]
	<i>Enterobacter aerogenes</i>	[40]
	<i>Flavobacterium spp</i>	[40]
	<i>Klebsiella ozaenae</i>	[40]
	<i>Klebsiella pneumoniae</i>	[40]
	<i>Proteus mirabilis</i>	[40]
	<i>Salmonella typhi</i>	[40]
	<i>Salmonella paratyphi</i>	[40]
<i>Serratia marcescens</i>	[40]	
<i>Shigella dysenteriae</i>	[40]	
<i>Helicobacter pylori</i>	[40]	
O. dictamnus	<i>Salmonella enteritidis</i>	[41]
	<i>Salmonella typhimurium</i>	[13]
	<i>Escherichia coli</i>	[13,20,33]
	<i>Enterobacter cloacae</i>	[33]
	<i>Klebsiella pneumoniae</i>	[33,42]
	<i>Pseudomonas aeruginosa</i>	[33,42]
	<i>Helicobacter pylori</i>	[41]
	<i>Acinetobacter hemolyticus</i>	[42]
	<i>Empedobacter brevis</i>	[42]
	<i>Erwinia carotovora</i>	[43]
<i>Clavibacter michiganensis</i>	[44]	
O. microphyllum	<i>Escherichia coli</i>	[20]
O. majorana	<i>Escherichia coli</i>	[25]
	<i>Pseudomonas aeruginosa</i>	[25]
	<i>Salmonella poona</i>	[25]
	<i>Helicobacter pylori</i>	[41]
O. acutidens	<i>Acinetobacter baumannii</i>	[34]
	<i>Acinetobacter lwoffii</i>	[34]
	<i>Brucella abortus</i>	[34]
	<i>Cedecea davisae</i>	[34]
	<i>Enterobacter cloacae</i>	[34]
	<i>Escherichia coli</i>	[34]
	<i>Klebsiella pneumoniae</i>	[34]
	<i>Morganella morganii</i>	[34]
	<i>Proteus vulgaris</i>	[34]
	<i>Pseudomonas aeruginosa</i>	[34]
	<i>Pseudomonas pseudoalkaligenes</i>	[34]
	<i>Salmonella choleraesuis arizonae</i>	[34]
	<i>Salmonella enteritidis</i>	[34]
	<i>Serratia plymuthica</i>	[34]
<i>Shigella sonnei</i>	[34]	
<i>Xanthomonas campestris</i>	[34]	
O. glandulosum	<i>Escherichia coli</i>	[35]
	<i>Klebsiella pneumoniae</i>	[35]
	<i>Pseudomonas aeruginosa</i>	[35]
O. minutiflorum	<i>Escherichia coli</i> O157:H7	[36]
	<i>Salmonella typhimurium</i>	[36]

Table 3: *Origanum* species having activity against fungi.

Origanum species	Fungi	Reference
O. syriacum	<i>Aspergillus fumigatus</i>	[3]
	<i>Aspergillus flavus</i>	[3]
	<i>Aspergillus niger</i>	[3]
	<i>Saccharomyces cerevisiae</i>	[10]
O. dictamnus	<i>Candida albicans</i>	[11]
	<i>Candida tropicalis</i>	[11]
	<i>Candida glabrata</i>	[11]
	<i>Botrytis cinerea</i>	[11]
	<i>Fusarium sp.</i>	[11]
	<i>Aspergillus niger</i>	[13]
	<i>Saccharomyces cerevisiae</i>	[13]
	<i>Penicillium digitatum</i>	[44]
O. acutidens	<i>Yarrowia lipolytica</i>	[45]
	<i>Candida albicans</i>	[34]
	<i>Alternaria solani</i>	[34]
	<i>Aspergillus flavus</i>	[34]
	<i>Aspergillus niger</i>	[34]
	<i>Aspergillus variegator</i>	[34]
	<i>Fusarium oxysporum</i>	[34]
	<i>Fusarium solani</i>	[34]
	<i>Microsporium canis</i>	[34]
	<i>Monilia fructicola</i>	[34]
	<i>Mortieraula alpina</i>	[34]
	<i>Penicillium spp.</i>	[34]
	<i>Rhizopus spp.</i>	[34]
	<i>Rhizoctonia solani</i>	[34]
	<i>Trichophyton rubrum</i>	[34]
	O. glandulosum	<i>Candida albicans</i>
<i>Candida tropicalis</i>		[35]
O. majorana	<i>Candida rugosa</i>	[46]
	<i>Debaryomyces hansenii</i>	[46]
	<i>Kluyveromyces marxianus</i>	[46]
	<i>Rhodotorula glutinis</i>	[46]
	<i>Rhodotorula minuta</i>	[46]
	<i>Saccharomyces cerevisiae</i>	[46]
	<i>Trichosporon cutaneum</i>	[46]
	<i>Yarrowia lipolytica</i>	[46]
	<i>Zygosaccharomyces rouxii</i>	[46]
O. vulgare	<i>Candida glabrata</i>	[47]
	<i>Microsporium canis</i>	[48]
	<i>Microsporium gypseum</i>	[48]
	<i>Trichophyton mentagrophytes</i>	[48]
	<i>Trichophyton erinacei</i>	[48]
	<i>Trichophyton terrestre</i>	[48]

Essential oil and extracts (aqueous, ethanolic and cyclohexane) of aerial parts of the plants have antioxidant activity. Also, infusion and decoction processes of these extracts have this activity [8,9,11,34,53,54].

It has been shown that the antioxidant activity improves endothelial function, has anti-inflammatory properties and stimulates the DNA repair mechanism [19]. Many studies have highlighted the importance of phenolic compounds and flavonoids of essential oils in the antioxidant activity [9,25,34,51,53,54]. Viuda-Martos, et al. [19] established that the antioxidant activity is possibly due to the presence of thymol in the essential oils, while in other studies this property was mainly attributed to the presence of carvacrol [13,34], rosmarinic acid and polyphenols [34]. Liolios, et al. [11] and Lukas, et al. [22] established that this ability is also due to other polar and non-polar phenolic compounds present in the extracts which are rich in phenolic derivatives.

Table 4: *Origanum* species with antioxidant activity.

Origanum species	Method of study	Reference
<i>O. ehrenbergii</i>	β-Carotene Bleaching Inhibition [CBI]	[7]
<i>O. syriacum</i>		[7]
<i>O. vulgare</i>		[9,25]
<i>O. majorana</i>		[25]
<i>O. acutidens</i>		[34]
<i>O. glandulosum</i>		[50]
<i>O. ehrenbergii</i>	2,2'-Diphenyl-1-Picrylhydrazyl [DPPH] radical-scavenging	[7]
<i>O. syriacum</i>		[7,10,19]
<i>O. vulgare</i>		[9,25,51]
<i>O. dictamnus</i>		[13,20]
<i>O. syriacum</i>		[16]
<i>O. minutiflorum</i>		[16]
<i>O.</i>		[20]
<i>mychrophylllum</i>		[20]
<i>O. libanoticum</i>		[25,52]
<i>O. majorana</i>		[34]
<i>O. acutidens</i>		[53]
<i>O. compactum</i>		
<i>O. vulgare</i>		Reducing Power [RP]
<i>O. syriacum</i>	[10]	
<i>O. majorana</i>	[52]	
<i>O. vulgare</i>	Ferric Reducing Antioxidant Capacity [FRAC] or Power [FRAP]	[9,51]
<i>O. syriacum</i>		[19]
<i>O. dictamnus</i>		[20]
<i>O.</i>		[20]
<i>mychrophylllum</i>		[20]
<i>O. libanoticum</i>		
<i>O. vulgare</i>	Inhibition of lipid peroxidation by TBARS assay	[9]
<i>O. majorana</i>		[52]
<i>O. syriacum</i>	Ascorbate-Iron [III]-Catalyzed Phospholipid Peroxidation.	[16]
<i>O. minutiflorum</i>		[16]
<i>O. syriacum</i>	Iron [III] to Iron [II] Reducing Activity	[16]
<i>O. minutiflorum</i>		[16]
<i>O. syriacum</i>	Iron [II] Chelation Activity	[16]
<i>O. minutiflorum</i>		[16]
<i>O. syriacum</i>	Site-Specific Hydroxyl Radical-Mediated 2-Deoxy-D-ribose Degradation	[16]
<i>O. minutiflorum</i>		[16]
<i>O. syriacum</i>	Nonsite-Specific Hydroxyl Radical-Mediated 2-Deoxy-D-ribose Degradation	[16]
<i>O. minutiflorum</i>		[16]
<i>O. syriacum</i>	Inhibition of Lipid Peroxidation LPI of Buffered Egg Yolk by TBARS assay	[19]
<i>O. syriacum</i>	Determination of Oxidative Stability of Fat [RANCIMAT Assay]	[19]
<i>O. syriacum</i>	Ferrous Ion Chelating [FIC] Ability	[19]
<i>O. vulgare</i>	Percent inhibition in linoleic acid system	[25]
<i>O. majorana</i>		[25]
<i>O. vulgare</i>	2,2'-Azino-Bis [3-Ethylbenzothiazoline]-6-Sulfonic acid [ABTS]; radical scavenging	[51]
<i>O. compactum</i>		[53]
<i>O. majorana</i>	Hydroxyl radical scavenging activity	[52]
<i>O. majorana</i>	Hydrogen peroxide scavenging activity	[52]

Therefore, we may assume that this activity may be the result of a combination of many substances being of high importance phenolics and flavonoids compounds.

Anti-inflammatory activity

Inflammation is a non-specific physiological response to environmental stressors such as infectious microorganisms, chemical agents, physical injuries and many diseases. This process seeks to destroy the offending agent and repair the damaged tissue or organ. Activated leucocytes, endothelial cells and macrophages produce pro-inflammatory cytokines such as Interleukin (IL) IL-1b, IL-6 and TNF-α (Tumor Necrosis Factor α) among others, and anti-inflammatory cytokines such as IL-10. These cells also produce pro-inflammatory enzymes such as inducible nitric oxide synthase and cyclooxygenase,

Table 5: *Origanum* species and anti-inflammatory activity performed.

Origanum species	Anti-inflammatory activity	Reference
<i>O. ehrenbergii</i>	Inhibit NO synthesis in murine RAW 264.7 cell line	[7]
<i>O. syriacum</i>	Inhibit NO synthesis in murine RAW 264.7 cell line	[7,21]
<i>O. vulgare spp Hirtum</i>	Inhibit NO synthesis in murine RAW 264.7 cell line	[21]
<i>O. vulgare</i>	Inhibit NO synthesis in murine RAW 264.7 cell line	[21]
	↓ IL-1b, IL-6 y TNF-α / ↑ IL-10	[55]
	↓ IL-1b, IL-6, GM-CSF and TNF-α in mice	[56]
	↓ T helper 17 cells / ↑ T helper 2 and T regulatory cells.	[57]

which increase the level of nitric oxide and prostaglandin E2 [55]. *Origanum* species act by inhibiting the secretion of pro-inflammatory cytokines and activating the expression of pro-inflammatory genes. Furthermore, this plant promotes the secretion of anti-inflammatory cytokines and Deactivates the expression of inflammatory genes. The anti-inflammatory activities studied for *Origanum* species are shown in Table 5.

In several species of *Origanum* (e.g. *O. vulgare*, *O. vulgare spp. Hirtum* and *O. syriacum*) rosmarinic, oleanolic and ursolic acids are present, and although their concentrations vary among these species, these acids could be responsible for the anti-inflammatory activity attribute to these plants. It was observed that the three acids previously mentioned decreased the expression of inducible nitric oxide synthase and cyclooxygenase genes, showing a strong anti-inflammatory activity. It seems that rosmarinic acid inhibits lipoxygenase and cyclooxygenase, while the mechanism of action of oleanolic and ursolic acids seems to focus on the involvement of one or more signaling pathways [21].

Methanolic extracts [57] and essential oils of *O. vulgare* have showed anti-inflammatory activity. The activity of the essential oil was studied in mouse models of gastritis induced stress and contact hypersensitivity [58]. Finally, Silva, et al. [59] has suggested that carvacrol present in the essential oil probably interferes with the release and/or synthesis of inflammatory mediators such as prostanoids.

Antitumor activity

Tumors, besides being very difficult to treat because of its enormous complexity and variability, are widespread and very serious diseases. It has been demonstrated antitumor activity on some species of *Origanum*, and furthermore, they have cytotoxic activity against several tumor cell lines. *Origanum* species with cytotoxic activity against different cell lines are presented in Table 6.

Arcila-Lozano, et al. [17] reported the antimutagenic and anticarcinogenic effect of several *Origanum sp.* Generally, essential oils [13,20,55,62,65], cyclohexane [11], dichloromethane extracts [18], ethyl acetate [53,63], ethanol [53,63,64] and aqueous extracts [60] have shown antitumor activity. This antitumoral activity has been attributed to various components, including carvacrol [13], ursolic acid [18], 4-terpineol [62] and betulinic acid [63]. Moreover, several studies have shown the presence of thymoquinone in the essential oil of *Origanum sp.* Thymoquinone is a molecule with antioxidant, anti-inflammatory and analgesic properties, promising as antitumor candidate [22].

Table 6: *Origanum* species and cell lines against which have cytotoxic activity.

Origanum species	Cell line	Reference
<i>O. dictamnus</i>	Deletes mutagenicity of Trp-P-2	[11]
	Breast cáncer, colon cancer and lung adenocarcinoma	[13]
	HepG2 [hepatic carcinoma]	[13,20]
	P388 y L-1210 [murine leukemia]	[18]
	NSCLC-N6 [bronchial epithelial tumor]	[18]
<i>O. mycophyllum</i>	LoVo [colon carcinoma]	[20]
	HepG2 [hepatic carcinoma]	[20]
<i>O. libanoticum</i>	LoVo [colon carcinoma]	[20]
	HepG2 [hepatic carcinoma]	[20]
<i>O. majorana</i>	LNCaP [prostate adedocarcinoma]	[25]
	NIH-3T3 [mouse fibroblasts]	[25]
	MCF-7 [breast adenocarcinoma]	[25,60]
	HeLa [cervical cancer]	[60]
<i>O. vulgare</i>	Jurkat [Acute lymphocytic leukemia]	[60]
	LNCaP [prostate adedocarcinoma]	[25]
	MCF-7 [breast adenocarcinoma]	[25,61]
	THP-1 [acute monocytic leukemia]	[55]
	HepG2 [hepatic carcinoma]	[61]
	NIH-3T3 [mouse fibroblasts]	[62]
<i>O. compactum</i>	HT-29 [colon adenocarcinoma]	[62]
	MCF-7 [breast adenocarcinoma]	[53,63]
<i>O. syriacum</i>	THP-1 [acute monocytic leukemia]	[64]
	PMBCs [Peripheral blood mononuclear cells]	[64]
<i>O. onites</i>	5RP7 [rat fibroblasts tumor]	[65]
	Rat adipose tissue endothelial cell migration	[65]

Antiparasitic activity

Parasitism is a process in which an organism (parasite) depends on the host and obtains a benefit from this. In many cases, the parasites cause disease in the hosts, which can be humans, animals and plants, so that this process has a lot of medical and agricultural importance. *Origanum* species with probe anti-parasitic activity are presented in Table 7.

Antiparasitic activity has been found in the essential oils [53,66,70], and acetyl [53,68] and in methanol extracts [69]. This activity may be due to the presence of phenolic compounds in the essential oil as thymol and carvacrol that interact with the permeability of the cytoplasmic cell membrane [66]. Antiparasitic activity is also due to the presence of terpenoids and flavonoids [68]. Interestingly, in some occasions, along with the antiparasitic activity there was an improvement of the innate immune and phagocytic activity in the host [70].

Antiviral activity

Viruses are acellular microscopic infectious agents that need the cells of other organisms (hosts) to multiply. Their hosts may be

Table 7: *Origanum* species and parasite that they affect.

Origanum species	Parasite	Reference
<i>O. vulgare</i>	Affects the development of malaria	[25]
	Coccidium of <i>Eimeria tenella</i>	[66]
	Coccidium of <i>Eimeria sp.</i>	[67]
<i>O. compactum</i>	<i>Plasmodium falciparum</i>	[53]
	<i>Schistosoma haematobium</i>	[68]
<i>O. syriacum</i>	Cysts and trophozoites of <i>Acanthamoeba castellanii</i>	[69]
<i>O. laevigatum</i>	Cysts and trophozoites of <i>Acanthamoeba castellanii</i>	[69]
<i>O. minutiflorum</i>	<i>Mixobolus sp.</i> [Bivalvulida/Platysporina]	[70]

humans, animals or plants. Many of these viruses cause disease, so the antiviral activity of *Origanum* species is also very important to medical and agricultural objectives. Works focus on *Origanum sp.* antiviral activity is presented in Table 8.

Hexane, dicholoromethane, methanol [34], aqueous extracts [71] and essential oils [72] have showed antiviral activity. In the study of Sökmen, et al. [34] this activity was ascribed to the presence of rosmarinic acid. Sanchez and Aznar [72] found that the essential oil slightly reduced the infectivity of HAV while thymol present in the essential oil reduced the infectivity of norovirus, murine norovirus and feline calicivirus.

Anticholinesterase activity

Cholinesterase's are enzymes that catalyze the hydrolysis of neurotransmitters in the synaptic space, necessary to allow the cholinergic neuron to return to its resting state after activation, avoiding overstimulation and damage to the neuron or muscle. Acetyl-cholinesterase catalyzes the cleavage of acetylcholine while butyryl-cholinesterase hydrolyzes butyrylcholine. Both enzymes play an important role in the central nervous system, and their inhibitors are used in Pharmacy. Malfunctions of these enzymes results in neurodegenerative diseases where oxidative stress plays an important role. *Origanum* species with tested anticholinesterase activity are detailed in Table 9.

Mossa and Nawwar [52] attributed this activity to the presence of terpenoids as 4-terpinenol γ -terpinene, α -terpinene, p-cymene and 1,8-cineol in the essential oils of these plants, while Loizzo, et al. [73] attributed this property to the presence of thymol and carvacrol. Newly, the combination of the compounds present in the essential oils of these plants seems to be responsible for this property and provides future applications of these plants in studies focus on the prevention of neurodegenerative disorders [7,52,55].

Antidiabetes activity

Diabetes is a growing global problem characterized by insulin deficiency and the subsequent presence of blood glucose (hyperglycemia). *O. vulgare* can delay the development of diabetic complications and correct metabolic abnormalities thanks to his hypoglycemic property [5,23]. Vujicic, et al. [57] observed that methanolic extracts of *Origanum* reduced diabetes and established a normal secretion of insulin in mice with diabetes induced by antioxidant, anti-inflammatory and antiapoptotic activity in β cells. Rosmarinic acid could be involved in this activity. In the study

Table 8: *Origanum* species and virus that they affect.

Origanum species	Virus	Reference
<i>O. acutidens</i>	Herpes simplex virus type 1 [HSV-1]	[34]
<i>O. vulgare</i>	Suid herpesvirus type 1 [SuHV-1]	[71]
	Hepatitis A virus [HAV]	[72]

Table 9: *Origanum* species and anticholinesterase activity present.

Origanum species	Affected enzyme	Reference
<i>O. ehrenbergii</i>	Acetyl-cholinesterasa	[7]
	Butyryl-cholinesterasa	[7]
<i>O. syriacum</i>	Acetyl-cholinesterasa	[7]
	Butyryl-cholinesterasa	[7]
<i>O. majorana</i>	Acetyl-cholinesterasa	[52]

developed by Lemhadri, et al. [74] the blood glucose levels decreased strongly in diabetic rats after treating them with *O. vulgare*. They observed that flavonoids can cause hypoglycemic effect acting separately or synergistically with other components.

Origanum Sp. as a Dietary Supplement

To the best of our knowledge, there is available few studies focus on the effects of dietary administration of *O. vulgare* on animal diets. However, the administration of these plants on diet to farm animals has yielded very good results. Dietary supplements of the essential oil from *O. vulgare* on chickens infected with *Eimeria tenella* produced weight gain of the chickens and reduced coccidiosis [66]. Similar results were observed in the study of Nosal, et al. [67], where this effect was observed against *Eimeria spp.* in rabbits (*Oryctolagus cuniculus f. domesticus*). It was also observed that the essential oil included in the diet increased weight and had positive effects against pathogenic organisms, as well as in the secretions of digestive and liver enzymes in poultry [75]. Also, there is a recent study in which the ethanolic extract of *O. vulgare* has been dietary administered to rainbow trout (*Oncorhynchus mykiss*) specimens and their innate immune response was improved [76]. In this study, one thousand and two hundreds rainbow trout were randomly allocated into two groups including placebo-treated group (control) and *O. vulgare* extract-treated group, each of three replicates. The fish were hand-fed once a day with each at a rate of 1% of feed weight in the first feeding for 8 weeks. At the end of every two weeks, blood samples were analyzed for some of hematological, biochemical and immunological parameters. Respiratory burst, phagocytosis and serum lysozyme of leucocytes was studied. The observed effects include enhancements in the innate immune response by increasing the levels of total serum proteins as albumin, globulin and lysozyme, as well as the respiratory burst and phagocytic activity of leucocytes [76].

Therefore, various studies using plants of the genus *Origanum* have yielded good results, showing that its inclusion in the diet increases the weight of the animals; helps fight infections of parasites and increase the innate immune response of the animals. These important effects on farmed animals are due to the existence in the essential oils of, among others, carvacrol, linalol, borneol, α -terpinene, γ -terpinene, α -pinene and β -pinene.

Chemical Composition

Because the primary responsibility for the properties of these plants is the essential oils, most of the chemical composition studies have focused on studying these essential oils. It has been found that although all species of the genus *Origanum* contain more or less the same components, chemical composition and concentrations of the components depends on many factors such as genotype, geographical origin, climate, type and soil composition, orientation the development of the plant, harvest time and culture conditions [3,4,15,17,77].

Essential oils of these species have alkaloids, flavonoids, phenolic and polyphenolic compounds [5,11,19,37,78]. The main components of the essential oil of these plants, in most cases, are two oxygen monoterpenes, carvacrol and thymol [3,12,78-80]. Thymol and carvacrol, which are isomeric phenols, are primarily responsible for the characteristics and properties of essential oils. The biosynthetic precursors of carvacrol and thymol are the γ -terpinene and p -cymene

[11]. Although thymol and carvacrol are present in most plants species and they are generally in high concentrations, other compounds are also commonly found in these plants, such as hydrocarbon monoterpenes γ -terpinene, p -cymene, α -terpinene, α -myrcene, α -pinene and sabinene hydrate, and oxygenated monoterpenes linalool, 4-terpineol and α -terpineol, and sesquiterpene β -caryophyllene [3,9-12,19,80].

The different *Origanum* species have chemotypes of carvacrol and thymol and their presence and concentration varies among them and also, within different plants from the same specie [3,7]. This is very important because these chemotypes influence biosynthetic pathways and therefore, the relative proportion of parent compounds and the characteristics of essential oils [3,81]. Furthermore, some other species have carvacrol and thymol chemotypes with a high content of two hydrocarbon monoterpenes, γ -terpinene or p -cymene, whereas other species have high values linalool, phenols, alcohols, ethers, aldehydes and ketones [77].

While thymol has antibacterial, anti-inflammatory, anti- protozoal, antioxidant, cytotoxic and piscicide activities, and is an allelopathic agent [82], carvacrol has anti-oxidant, anti-microbial, antitumor, antimutagenic, antigenotoxic, analgesic, anti-spasmodic, anti-inflammatory, angiogenic, antiparasitic, antiplatelet, inhibiting pain, elastase, insecticide, antihepatotoxic, neuroprotective and hepatoprotective activities [83,84].

Conclusion

Plants of the genus *Origanum* have many and varied benefits that give them a great importance for being used in foods, as well as in pharmaceutical, medical and agricultural purposes. However, till present most of these beneficial effects have been seen almost exclusively in in-vitro studies, using human and mouse cell lines. It would be very interesting to perform more in vivo studies to know if the effects are comparable or not with the available in vitro effects. Until now, some in vivo studies carried out in animals have demonstrated their beneficial properties, which could make these plants good candidates for being included in animal feed, in as much that the inclusion of plants of genus *Origanum* in the diet of animals increases the weight of the animals, helps fight infections and improve the immune status.

Both, essential oils or extracts from aerial parts of the plants could be used for in vivo studies because the combination of the compounds present in them seems to be responsible of the activity of these plants, all acting synergistically.

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References

1. Salvador Cañigueral, Eduardo Dellacassa and Arnaldo L Bandoni. Plantas Medicinales y Fitoterapia: ¿Indicadores de Dependencia o Factores de Desarrollo? Acta Farmaceutica Bonaerense. 2003; 22: 265-278.
2. Nidal Amin Jaradat. Review of the taxonomy, ethnobotany, phytochemistry, phytotherapy and phytotoxicity of germander plant (*Teucrium polium* L.). Asian Journal of Pharmaceutical and Clinical Research. 2015; 8.

3. Abd El-Nasser El Gendy, Michele Leonardi, Linda Mugnaini, Fabrizio Bertelloni, Valentina VE, Simona Nardoni, et al. Chemical composition and antimicrobial activity of essential oil of wild and cultivated *Origanum syriacum* plants grown in Sinai, Egypt. *Industrial Crops and Products*. 2015; 67: 201-207.
4. Rodriguez-Garcia I, Silva-Espinoza BA, Ortega-Ramirez LA, Leyva JM, Siddiqui MW, Cruz-Valenzuela MR, et al. Oregano Essential Oil as an Antimicrobial and Antioxidant Additive in Food Products. *Critical Reviews in Food Science and Nutrition*. 2015; 12.
5. Shayista Chishty, Zahoor A. Kaloo and Phalestine Sultan. Medicinal importance of genus *Origanum*: A review. *Journal of Pharmacognosy and Phytotherapy*. 2013; 5: 170-177.
6. Letswaart JH. A taxonomic revision of the genus *Origanum* (Labiatae), Leiden Botanical series, Leiden University Press, The Hague, Leiden. 1980; 4.
7. Loizzo MR, Menichini F, Conforti F, Tundis R, Bonesi M, Saab AM, et al. Chemical analysis, antioxidant, anti-inflammatory and anticholinesterase activities of *Origanum ehrenbergii* Boiss and *Origanum syriacum* L. essential oils. *Food Chemistry*. 2009; 117: 174-180.
8. Barros L, Heleno SA, Carvalho AM and Ferreira CFR. Lamiaceae often used in Portuguese folk medicine as a source of powerful antioxidants: vitamins and phenolics. *Food Science and Technology*. 2010; 43: 544-550.
9. Martins N, Barros L, Santos-Buelga C, Henriques M, Silva S, Ferreira IC. Decoction, infusion and hydroalcoholic extract of *Origanum vulgare* L.: Different performances regarding bioactivity and phenolic compounds. *Food Chemistry*. 2014; 158: 73-80.
10. Mehmet Hakki Alma, Ahmet Mavi, Ali Yildirim, Metin Digrak, and Toshifumi Hirata. Screening Chemical Composition and in Vitro Antioxidant and Antimicrobial Activities of the Essential Oils from *Origanum syriacum* L. Growing in Turkey. *Biological and Pharmaceutical Bulletin*. 2003; 26: 1725-1729.
11. Christos Liolios, Konstantia Graikou, Eleni Skaltsa and Ioanna Chinou. Dittany of Crete: A botanical and ethnopharmacological review. *Journal of ethnopharmacology*. 2010; 131: 229-241.
12. Sivropoulou A, Papanikolaou E, Nikolaou C, Kokkini S, Lanaras T, Arsenakis M, et al. Antimicrobial and Cytotoxic Activities of *Origanum* Essential Oil. *Journal of Agricultural and Food Chemistry*. 1996; 44: 1202-1205.
13. Mitropoulou G, Fitsiou E, Stavropoulou E, Papavassilopoulou E, Vamvakias M, Pappa A, et al. Composition, antimicrobial, antioxidant, and antiproliferative activity of *Origanum dictamnus* (dittany) essential oil. *Microbial Ecology in Health Disease*. 2015; 6: 26543.
14. Salud Pérez G, Miguel Zavala S, Lucina Arias G and Miguel Ramos L. Anti-inflammatory Activity of Some Essential Oils. *Journal of essential oil research*. 2011; 23: 38-44.
15. Burt S. Essential oils: their antibacterial properties and potential applications in foods—a review. *International Journal of Food Microbiology*. 2004; 94: 223-253.
16. HJ Damien Dorman, Oliver Bachmayer, Müberra Kosar, Raimo Hiltunen. Antioxidant Properties of Aqueous Extracts from Selected Lamiaceae Species Grown in Turkey. *Journal of Agricultural and Food Chemistry*. 2004; 52: 762-770.
17. Arcila-Lozano CC, Loarca-Piña G, Lecona-Urbe S and González de Mejía E. Oregano: properties, composition and biological activity. *Archivos Latinoamericanos de Nutrición*. 2004; 54:100-111.
18. Chinou I, Liolios C, Moreau D and Roussakis C. Cytotoxic activity of *Origanum dictamnus*. *Phytotherapy Research*. 2007; 78: 342-344.
19. Manuel Viuda-Martos, Abd El-Nasser G-S El Gendy, Esther Sendra, Juana Fernández-López, KA Abd El Razik, Omer EA, et al. Chemical Composition and Antioxidant and Anti-Listeria Activities of Essential Oils Obtained from Some Egyptian Plants. *Journal of Agricultural and Food Chemistry*. 2010; 58: 9063-9070.
20. Mariangela Marrelli, Filomena Confortia, Carmen Formisano, Daniela Rigano, Nelly Apostolides Arnold, Menichini F, et al. Composition, antibacterial, antioxidant and antiproliferative activities of essential oils from three *Origanum* species growing wild in Lebanon and Greece. *Natural Product Research*. 2015: 1-5.
21. Diandian Shen, Min-Hsiung Pan, Qing-Li Wu, Chung-Heon Park, H. Rodolfo Julian, Ho CT, et al. LC-MS Method for the Simultaneous Quantitation of the Anti-inflammatory Constituents in Oregano (*Origanum* Species). *Journal of Agricultural and Food Chemistry*. 2010; 58: 7119-7125.
22. Brigitte Lukas, Corinna Schmiderer, Chlodwig Franz and Johannes Novak. Composition of Essential Oil Compounds from Different Syrian Populations of *Origanum syriacum* L. (Lamiaceae). *Journal of Agricultural and Food Chemistry*. 2009; 57: 1362-1365.
23. A Saravana Kumar, S Kavimani, and KN Jayaveera. A review on medicinal plants with potential antidiabetic activity. *International Journal of Phytopharmacology*. 2011; 2: 53-60.
24. Saeed S and Tariq P. Antibacterial activity of oregano (*Origanum vulgare* Linn.) against gram positive bacteria. *Pakistan Journal of Pharmaceutical Science*. 2009; 22: 421-424.
25. Abdullah I. Hussain, Farooq Anwar, Shazia Rasheed, Poonam S. Nigam, Omar Janneh, Satyajit D Sarker. Composition, antioxidant and chemotherapeutic properties of the essential oils from two *Origanum* species growing in Pakistan. *Revista Brasileira de Farmacognosia*. 2011; 21: 943-952.
26. Juliana de Lima Marques, Lisiane Martins Volcão, Graciele Daiana Funck, Isabela Schneid Kroning, Wladimir Padilha da Silva, Ângela Maria Fiorentini, et al. Antimicrobial activity of essential oils of *Origanum vulgare* L. and *Origanum majorana* L. against *Staphylococcus aureus* isolated from poultry meat. *Industrial Crops and Products*. 2015; 77: 444-450.
27. Melo AD, Amaral AF, Schaefer G, Luciano FB, de Andrade C, Costa LB, et al. Antimicrobial effect against different bacterial strains and bacterial adaptation to essential oils used as feed additives. *Canadian Journal Veterinary Research*. 2015; 79: 285-289.
28. Teixeira B, Marques A, Ramos C, Serrano C, Matos O, Neng NR, et al. Chemical composition and bioactivity of different oregano (*Origanum vulgare*) extracts and essential oil. *Journal of the Science of Food and Agriculture*. 2013; 93: 2707-2714.
29. Suzana ID, Katarina RM, Dusan GA, Mirjana RMS and Dusan ZM. A study of the synergistic antilisterial effects of a sublethal dose of lactic acid and essential oils from *Thymus vulgaris* L., *Rosmarinus officinalis* L. and *Origanum vulgare* L. *Food Chemistry*. 2007; 104: 774-782.
30. Elvia Hernández-Hernández, Carlos Regalado-González, Pedro Vázquez-Landaverde, Isabel Guerrero-Legarreta, and Blanca E. García-Almendárez. Microencapsulation, Chemical Characterization, and Antimicrobial Activity of Mexican (*Lippia graveolens* H.B.K.) and European (*Origanum vulgare* L.) Oregano Essential Oils. *Scientific World Journal*. 2014; 641814.
31. Economakis C, Demetzos C, Anastassaki T, Papazoglou V, Gazouli M, Loukis A, et al. Volatile constituents of bracts and leaves of wild and cultivated *Origanum dictamnus*. *Planta Medica*. 1999; 65: 189-191.
32. Economakis CD, Skaltsa H, Demetzos C, Sokovic M and Thanos CA. Effect of phosphorus concentration of the nutrient solution on the volatile constituents of leaves and bracts of *Origanum dictamnus*. *Journal of Agricultural and Food Chemistry*. 2002; 50: 6276-6280.
33. Liolios CC, Gortzi O, Lalas S, Tsaknis J and Chinou I. Liposomal incorporation of carvacrol and thymol isolated from the essential oil of *Origanum dictamnus* L. and in vitro antimicrobial activity. *Food Chemistry*. 2009; 112: 77-83.
34. Sökmen M, Serkedjieva J, Daferera D, Gulluce M, Polissiou M, Tepe B, et al. In vitro antioxidant, antimicrobial, and antiviral activities of the essential oil and various extracts from herbal parts and callus cultures of *Origanum acutidens*. *Journal of Agricultural and Food Chemistry*. 2004; 52: 3309-3312.
35. K Mechergui, W Jaouadi, JA Coelho, MC Serra and ML Khouja. Biological

- activities and oil properties of *Origanum glandulosum* Desf: A Review. *Phytothérapie*. 2015.
36. Dadalioglu I and Evrendilek GA. Chemical compositions and antibacterial effects of essential oils of Turkish oregano (*Origanum minutiflorum*), bay laurel (*Laurus nobilis*), Spanish lavender (*Lavandula stoechas* L), and fennel (*Foeniculum vulgare*) on common foodborne pathogens. *Journal of Agricultural and Food Chemistry*. 2004; 52: 8255-8260.
37. Darwish RM and Aburjai TA. Effect of ethnomedicinal plants used in folklore medicine in Jordan as antibiotic resistant inhibitors on *Escherichia coli*. *BioMed Central Complementary and Alternative Medicine*. 2010; 10: 9.
38. Ayman Al-Mariri and Maren Safi. In Vitro Antibacterial Activity of Several Plant Extracts and Oils against Some Gram-Negative Bacteria. *Iran Journal of Medical Sciences*. 2014; 39: 36-43.
39. Ayman Al-Mariri and Mazen Safi. The Antibacterial Activity of Selected Labiatae (Lamiaceae) Essential Oils against *Brucella melitensis*. *Iran Journal of Medical Sciences*. 2013; 38: 44-50.
40. Nazia Masood Ahmed Chaudhry, Sabahat Saeed and Perween Tariq. Antibacterial effects of oregano (*Origanum vulgare*) against gram negative bacilli. *Pakistan Journal of Botany*. 2007; 39: 609-613.
41. Stamatis G, Kyriazopoulos P, Golegou S, Basayiannis A, Skaltsas S, Skaltsa H. In vitro anti-*Helicobacter pylori* activity of Greek herbal medicines. *Journal of Ethnopharmacology*. 2003; 88: 175-179.
42. Chatzopoulou A, Karioti A, Gousiadou Ch, Lax Vivancos V, Kyriazopoulos P, Golegou S et al. Depsides and other polar constituents from *Origanum dictamnus* L. and their in vitro antimicrobial activity in clinical strains. *Journal of Agricultural and Food Chemistry*. 2010; 58: 6064-6068.
43. Vokou D, Varelzidou S and Katinakis P. Effects of aromatic plants on potato storage-sprouts suppression and antimicrobial activity. *Agriculture Ecosystems & Environment*. 1993; 47: 223-235.
44. Daferera DJ, Ziogas BN and Polissiou MG. GC-MS analysis of essential oils from some greek aromatic plants and their fungi toxicity on *Penicillium digitatum*. *Journal of Agricultural and Food Chemistry*. 2000; 48: 2576-2581.
45. Karanika MS, Komaitis M and Aggelis G. Effect of aqueous extracts of some plants of Lamiaceae family on the growth of *Yarrowia lipolytica*. *International Journal of Food Microbiology*. 2001; 64: 175-181.
46. Alina Kunicka-Styczyńska. Activity of essential oils against food-spoiling yeast. A review. *Flavour and Fragrance Journal*. 2011; 26: 326-328.
47. Soares IH, Loreto ÉS, Rossato L, Mario DN, Venturini TP et al. In vitro activity of essential oils extracted from condiments against fluconazole-resistant and -sensitive *Candida glabrata*. *Journal de Mycologie Medicale*. 2015; 25: 213-217.
48. Nardoni S, Giovanelli S, Pistelli L, Mugnaini L, Profili G, Pisseri F, et al. In Vitro Activity of Twenty Commercially Available, Plant-Derived Essential Oils against Selected Dermatophyte Species. *Nature Product Communication*. 2015; 10: 1473-1478.
49. Portillo-Ruiz MC, Sánchez RA, Ramos SV, Muñoz JV, Nevárez-Moorillón GV. Antifungal effect of Mexican oregano (*Lippia berlandieri* Schauer) essential oil on a wheat flour-based medium. *Journal of Food Science*. 2012; 77: M441-M445.
50. Belhattab R, Larous LA, Figueiredo C, Pedro AG Santos, José G Barroso, Luis G Pedro. *Origanum glandulosum* Desf. grown wild in Algeria: essential oil composition and glycosidic bound volatiles. *Flavour and Fragrance Journal*. 2005; 20: 209-212.
51. Valeria Velasco and Pamela Williams. Improving meat quality through natural antioxidants. *Chilean Journal of Agricultural Research*. 2011; 71.
52. ATH Mossa and GAM Nawwar. Free radical scavenging and antiacetylcholinesterase activities of *Origanum majorana* L. essential oil. *Human and Experimental Toxicology*. 2011; 30: 1501-1513.
53. Fatiha El Babili, Jalloul Bouajila, Jean Pierre Souchard, Cédric Bertrand, Florian Bellvert, Fouraste I, et al. Oregano: Chemical Analysis and Evaluation of Its Antimalarial, Antioxidant, and Cytotoxic Activities. *Journal of Food Science*. 2011; 76: c512-518.
54. Kaurinovic B, Popovic M, Vlasisavljevic S and Trivic S. Antioxidant capacity of *Ocimum basilicum* L. and *Origanum vulgare* L. extracts. *Molecules*. 2011; 16: 7401-7414.
55. A Ocaña-Fuentes, E Arranz-Gutiérrez, FJ Señorans and G Reglero. Supercritical fluid extraction of oregano (*Origanum vulgare*) essentials oils: Anti-inflammatory properties based on cytokine response on THP-1 macrophages. *Food and Chemical Toxicology*. 2010; 48: 1568-1575.
56. Bukovska A, Cikos S, Juhas S, Ilkova G, Rehak P, Koppel J. Effects of a combination of thyme and oregano essential oils on TNBS-induced colitis in mice. *Mediators of Inflammation*. 2007; 2007: 23296.
57. Vujicic M, Nikolic I, Kontogianni VG, Saksida T, Charisiadis P, Orescanin-Dusic Z, et al. Methanolic extract of *Origanum vulgare* ameliorates type 1 diabetes through antioxidant, anti-inflammatory and anti-apoptotic activity. *British Journal of Nutrition*. 2015; 113: 770-782.
58. Yoshino K, Higashi N and Koga K. Antioxidant and anti-inflammatory activities of oregano extract. *Journal of Health Sciences*. 2006; 52: 169-173.
59. Silva FV, Guimaraes AG, Silva ER, Sousa-Neto BP, Machado FD, Orescanin-Dusic Z, et al. Anti-inflammatory and anti-ulcer activities of carvacrol, a monoterpene present in the essential oil of oregano. *Journal of Medicinal Food*. 2012; 15: 984-991.
60. Elansary HO and Mahmoud EA. Egyptian herbal tea infusions' antioxidants and their antiproliferative and cytotoxic activities against cancer cells. *Natural Product Research*. 2015; 29: 474-479.
61. Marrelli M, Cristaldi B, Menichini F and Conforti F. Inhibitory effects of wild dietary plants on lipid peroxidation and on the proliferation of human cancer cells. *Food Chemical Toxicology*. 2015; 86: 16-24.
62. Begnini KR, Nedel F, Lund RG, Carvalho PH, Rodrigues MR, Beira FT, et al. Composition and antiproliferative effect of essential oil of *Origanum vulgare* against tumor cell lines. *Journal of Medicinal Food*. 2014; 17: 1129-1133.
63. Wahid Chaouki, David Y. Leger, Jamila Eljastimi, Jean-Louis Beneytout and Mohamed Hmamouchi. Antiproliferative effect of extracts from *Aristolochia baetica* and *Origanum compactum* on human breast cancer cell line MCF-7. *Pharmaceutical Biology*. 2010; 48: 269-274.
64. Basim M Ayeshe, Abdalla A Abed and Doa'a M Faris. In vitro inhibition of human leukemia THP-1 cells by *Origanum syriacum* L. and *Thymus vulgaris* L. extracts. *BioMed Central Research Notes*. 2014; 7: 612.
65. Rakibe Beklem Bostancıoğlu, Mine Kürkcüoğlu, Kemal Hüsnü Can Baser and Ayşe Tansu Koparal. Assessment of anti-angiogenic and anti-tumoral potentials of *Origanum onites* L. essential oil. *Food and Chemical Toxicology*. 2012; 50: 2002-2008.
66. I Giannenas, P Florou-Paneri, M Papazahariadou, E Christaki, NA Botsoglou, Spais AB. Effect of dietary supplementation with oregano essential oil on performance of broilers after experimental infection with *Eimeria Tenella*. *Archives of Animal Nutrition*. 2003; 57: 99-106.
67. Paweł Nosal, Dorota Kowalska, Paweł Bielański, Jerzy Kowal and Sławomir Kornaś. Herbal formulations as feed additives in the course of rabbit subclinical coccidiosis. *Annals of Parasitology*. 2014; 60: 65-69.
68. Lahlou M. Potential of *Origanum compactum* as a cercaricide in Morocco. *Annals of Tropical Medicine and Parasitology*. 2002; 96: 587-593.
69. Serpil Degerli, Bektas Tepe, Ali Celiksoz, Seyda Berk and Erdogan Malatyali. In vitro amoebicidal activity of *Origanum syriacum* and *Origanum laevigatum* on *Acanthamoeba castellanii* cysts and trophozoites. *Experimental Parasitology*. 2012; 131: 20-24.
70. Karagouni E, Athanassopoulou F, Lytra A, Komis C and Dotsika E. Antiparasitic and immunomodulatory effect of innovative treatments against *Myxobolus* sp. infection in *Diplodus puntazzo*. *Veterinary Parasitology*. 2005; 134: 215-228.

71. MJB Fernandes, AV Barros, MS Melo and IC Simoni. Screening of Brazilian plants for antiviral activity against animal herpes viruses. *Journal of Medicinal Plants Research*. 2012; 6: 2261-2265.
72. Sánchez G and Aznar R. Evaluation of Natural Compounds of Plant Origin for Inactivation of Enteric Viruses. *Food and Environmental Virology*. 2015.
73. Loizzo MR, Tundis R, Menichini F, and Menichini F. Natural products and their derivatives as cholinesterase inhibitors in the treatment of neurodegenerative disorders: An update. *Current Medicinal Chemistry*. 2008; 15: 1209-1228.
74. Lemhadri A, Zeggwagh NA, Maghrani M, Jouad H and Eddouks M. Anti-hyperglycaemic activity of the aqueous extract of *Origanum vulgare* growing wild in Tafilalet region. *Journal of Ethnopharmacology*. 2004; 92: 251-256.
75. Lenuța Fotea, Elena Costăchescu, G. Hoha and Doina Leonte. The effect of oregano essential oil (*Origanum vulgare*) on broiler performance. *Lucrări Științifice, Seria Zootehnie*. 2015; 53: 491-494.
76. Masoud Haghighi and Mostafa Sharif Rohani. Non-Specific Immune Responses and Hematological Parameters of Rainbow Trout (*Oncorhynchus mykiss*) Fed with *Origanum vulgare* Extract Diets. *American Advances Journal of Biological Sciences*. 2015; 1: 1-9.
77. Russo M, Galletti GC, Bocchini P and Carnacini A. Essential oil chemical composition of wild populations of Italian Oregano spice (*Origanum vulgare* ssp. *hirtum* (Link) Ietswaart): a preliminary evaluation of their use in chemotaxonomy by cluster analysis 1. *Inflorescences. Journal of Agricultural and Food Chemistry*. 1998; 46: 3741-3746.
78. Bakkali F, Averbeck S, Averbeck D, Idaomar M. Biological effects of essential oils. A review. *Food and Chemical Toxicology*. 2008; 46: 446-475.
79. Sarer E, Scheffler JJ and Baerheim SA. Monoterpenes of the essential oil of *Origanum majorana*. *Planta Medica*. 1982; 46: 236-239.
80. Skoula M and Harborne JB. The Genera *Origanum* and *Lippia*. *Medicinal and Aromatic Plants-Industrial Profiles. Taxonomy and Chemistry*. 2002; 25: 67-108.
81. Viuda-Martos M, Mohamady MA, Fernandez-Lopez J, Abd El Razik KA, Omer EA, Jose Angel Pérez-Álvarez. In vitro antioxidant and antibacterial activities of essential oils obtained from Egyptian aromatic plants. *Food Control*. 2011; 22: 1715-1722.
82. Armando Talavera-Alemán, Gabriela Rodríguez-García, Yliana López, Hugo A García-Gutiérrez, J Martín Torres-Valencia, Rosa E del Río, et al. Systematic evaluation of thymol derivatives possessing stereogenic or prostereogenic centers. *Phytochemistry Reviews*. 2015; 10: 9412-9416.
83. Baser KH. Biological and pharmacological activities of carvacrol and carvacrol bearing essential oils. *Current Pharmaceutical Design*. 2008; 14: 3106-3119.
84. Mendel Friedman. Chemistry and Multibeneficial Bioactivities of Carvacrol (4-Isopropyl-2-methylphenol), a Component of Essential Oils Produced by Aromatic Plants and Spices. *Journal of Agricultural and Food Chemistry*. 2014; 62: 7652-7670.