Peripheral Smear Validity along with Automated Analyzer Regarding Platelet Count

Kanmani Devi M and Arun Kumar T*
Karpagam Faculty of Medical Sciences and Research and Hospital

Introduction

Automation in laboratory diagnostics has revolutionized the patient care practices in the past few years. Automated counts are being widely accepted in diagnostic field due to various advantages. Despite the sophistication of present day instruments there is still need to depend on manual techniques for primary calibration especially in case of platelet count. Automated analyzer had pitfalls due to mimickers of platelet like particles which validated the manual count in peripheral smear regarding platelet count by leishman’s stain.

Evaluation of platelet counts is crucial for certain conditions like dengue fever. Platelet counts are considered the most valuable parameter to assess the intensity of the illness and also to monitor the progression of the illness and response to treatment. In such critical situations, manual counting of platelets are considered more reliable compared to automated analyzers. For many years Brecher and Cronkite method was considered the ideal method due to its reliability, accuracy, time and cost. [3] In situations involving daily clinical practice, the need for accuracy and reliability of platelet counts are far more higher. Therefore, this study was done to evaluate the validity of manual counting and automated autoanalyzer counts in peripheral smear for platelet counts.

Material and Methods

Study setting

This study was carried out as a cross sectional study in our tertiary care hospital for a period of three months from April to June 2018.

Study samples

All the samples requested for peripheral smear during the study period were included. Overall, 1200 venous samples were selected with EDTA as the anticoagulant in vacutainers. Among the...
routine 100 to 120 samples/day, platelet counts < 100,000 lakh/cu.mm were taken up for the study. A total of 500 samples were subjected for study.

**Data collection tools**

Samples were processed by Symex 5000 autoanalyzer and compared with manual platelet count by leishman’s stain. Data regarding the background characteristics like age, sex of the patients were noted from the laboratory records. All manual platelet counts were analyzed using standard hematological method described by Dacie and Lewis, while the automated analyzers were done following the manufacturers guidelines. All samples were analyzed within 30 minutes.

**Ethical approval**

Approval was obtained from the Institutional Ethics Committee prior to the commencement of the study.

**Data analysis**

Data was entered and analyzed using Microsoft Excel spreadsheet for Windows 10. The results were expressed as percentages.

**Results**

About 500 samples with platelet counts < 100,000 lakh/cu.mm were analyzed. Among the study samples, 241 were inpatients and 259 were out patient samples. It was observed that in 384/500 (76.8%) haematology analyzed correlated with peripheral smear review. In about 116 (23%) there was no correlation between the manual counting and automated counting. Moreover, there were RBCs and large platelets in peripheral smear counted as platelets by autoanalyzer (Table 1).

**Discussion**

In the study we did not obtain significant variation in results related with age and sex, male or female, younger or older individuals by counting platelets manually and in automatic analyzer. When platelets are activated they become spherical with hypogranular cytoplasm and release small particles. These particles mostly give erroneous results regarding automation but can be rectified with such suspected cases using manual smear examination method. Limitation of technology, automated platelet count can be inaccurate even at normal or high platelet ranges owing to the presence of substantial amount of interfering particles including WBC’s fragments, RBC’s fragments, bacteria, lipid droplets and bacteria etc.,. But in case of neonatal thrombocytopenia, i.e. due to placental insufficiency, fetal hypoxia, sepsis, Necrotizing enterocolitis, viral infections includes cytomegalovirus, Rubella virus, staphylococcus infection, Escherichia Coli, etc., platelet count has to be done manually for accurate diagnosis. Limitation of technology, automated platelet count can be inaccurate even at normal or high platelet ranges owing to the presence of substantial amount of interfering particles including WBC’s fragments, RBC’s fragments, bacteria, lipid droplets and bacteria etc.,.

In a study done by De la Salle BJ et al, around 67% of the automated analyzer results were found to be overestimated. Moreover, statistically significant differences in platelet counts were observed in 16.5% of the cases.[4] There are however newer modalities using two dimensional counting systems with the automated analyzer, employed by the ADVIA counters which check for accuracy in differentiating platelets with non platelet particles, better compared to one dimensional counting systems.[5] In a study done by Bakhubaira et al, there was a positive correlation observed between manual and automated analyzers. This correlation was found to be statistically significant. This study also observed a difference in the mean platelet counts between both the techniques [3].

**Conclusion**

Our study concludes there was significant correlation between automated and manual counts. But this was not applicable for very high or low platelet counts. As platelet count estimation is very important element of the diagnostic and treatment disorder. The accuracy depends on the instrument bias and activated platelet components whereas in manual count this was eliminated. The examination of peripheral blood smear using leishman’s stain did not reveal any interference with non platelet particles. Peripheral smear examination along with automated analyzer valid in case of high or low value platelet counts.

**References**


**Citation:** Devi KM and Kumar AT. Peripheral Smear Validity along with Automated Analyzer Regarding Platelet Count. SM J Hematol Oncol. 2018; 3(1): 1010s1.