



Malignant Gastrointestinal Stromal Tumor: Case Report with Uncommon Presentation and Review of the Literature

Christian Green*, Nicholas Palagonia, Petr Stastka, Andrea Cevallos, Elizabeth O'Grady, Zachary Elder, and Mohamed Aziz

Department of Pathology- American University of the Caribbean, School of Medicine, USA

Abstract

Gastrointestinal stromal tumors (GISTs) are rare, making up less than 1% of all gastrointestinal tumors. Each year, approximately 4,000 to 6,000 adults in the United States will be diagnosed with a GIST. However, they are common mesenchymal tumors accounting for 80% of gastrointestinal mesenchymal tumors, as well as comprising 3-5% of all sarcomas and 1% of all gastrointestinal neoplasms. GISTs can occur along any location in the GI tract, but most GISTs are found in the stomach (60%), as well as the jejunum and ileum of the small intestine (20-30%). This case describes a 49-year-old male who presents with a malignant recurrent GIST two years after resection of a primary GIST. GISTs represent a problematic subset of gastrointestinal neoplasms due to the variability of immunohistochemistry and cytogenetics of the tumors across patients. GISTs should always be included in the differential diagnosis of presentation of a gastrointestinal mass. Early detection combined with complete resection of the mass, administration of tyrosine kinase inhibitors, as well as observation for possible recurrence is considered the current standard of care in patients with GIST.

KEYWORDS: Gastrointestinal, Malignant, immunohistochemistry, Imatinib, Mutation

ABBREVIATION

GIST: Gastrointestinal stromal tumors, **EGIST:** Extra-gastrointestinal gastrointestinal stromal tumors, **IHC:** Immunohistochemistry, **FNA:** Fina needle aspiration, **EUS-FNA:** Endoscopic ultrasonography-guided fine-needle aspiration

INTRODUCTION

Gastrointestinal stromal tumors (GIST) account for 80% of gastrointestinal mesenchymal tumors, and 1-3% of all gastrointestinal neoplasms [1,2]. GISTs can occur along any location in the GI tract, but most GISTs are found in the stomach (60%), as well as the jejunum and ileum of the small intestine (20-30%) [3]. However, GISTs are not limited to the GI tract; some have been discovered in the pancreas and gallbladder and are known as extra-gastrointestinal GIST (EGIST), though these are considered exceptional or metastasis from malignant GIST tumors located in the GI tract [4]. GISTs were considered rare tumors until 2000, often being misclassified as leiomyomas, leiomyoblastomas, or

leiomyosarcomas [1]. Advances in immunohistochemical studies and the discovery of the activating mutation of the C-kit tyrosine kinase allowed diagnosing GISTs commonplace. The current understanding of GISTs is that they arise from interstitial cells of Cajal, which are derived from mesoderm and serve as the pacemaker cells for the gastrointestinal tract [5,6].

Although GISTs are the most common mesenchymal tumors of the GI tract, mesenchymal tumors only constitute 1% of all primary gastrointestinal cancers [7]. GISTs are predominantly found in middle-aged or older individuals, with the mean age of diagnosis being at 64 years of age [8]. Although the majority of GISTs are sporadic, 5% of patients diagnosed with primary familial GIST syndrome, neurofibromatosis type 1, and Carney-Stratakis syndrome carry a predisposition [9]. GISTs are rare in pediatric patients, but are linked to mutations in succinate dehydrogenase, as well as Carney-Stratakis syndrome [10]. Interestingly, the majority of pediatric GISTs have distinct characteristics from adult GISTs, including the absence of a mutation in C-kit tyrosine kinase in 85% of cases [11]. GISTs, can be presented as a benign or as a malignant tumor, however, some investigators suggested that "benign" or "malignant" classification is not necessarily applied as they are not useful in patient management; instead, classification is focused on rate of recurrence and risk of metastasis [12-14].

GISTs represent a problematic subset of gastrointestinal neoplasms due to diagnostic challenges they present. Most GISTs (70%) show spindle-cell morphology, although a significant amount of GISTs show pure epithelioid morphology (20%) or mixed cellularity (10%) [2,12]. Therefore, spindle cell tumors, epithelioid cell tumors and mixed cell tumors are subject to differential diagnoses of GIST. In contrast to spindle-cell type GISTs that stain strongly for C-kit, epithelioid-cell type GISTs

Submitted: 09 February, 2021 | **Accepted:** 02 March, 2021 | **Published:** 05 March, 2021

***Corresponding author(s):** Christian Green, Department of Pathology, American University of the Caribbean School of Medicine, 1 University Drive at Jordan Road Cupecoy, St. Maarten, Tel: +16013071393; Email: christiangreen@students.aucmed.edu

Copyright: © 2021 Kahler K, et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Citation: Green C, Palagonia N, Stastka P, Cevallos A, O'Grady E, et al. (2021) Malignant Gastrointestinal Stromal Tumor: Case Report with Uncommon Presentation and Review of the Literature. SM J Sarcoma Res 5: 5.



have different staining patterns of C-kit. Pediatric GISTs share different characteristics and mutations than familial and adult GISTs, making the differential even more broad [9-11].

There are no effective preventative measures for GIST, as it is linked with advanced age and certain inherited conditions. Early detection combined with complete resection of the mass, administration of tyrosine kinase inhibitors, as well as observation for possible recurrence is considered the current standard of care in patients with GIST [15].

CASE PRESENTATION

A 49-year-old man with history of pre-gastric Gastrointestinal Stromal Tumor (GIST) presented two years later with a large abdominal mass. The earlier tumor was surgically removed with clear surgical margins and no post-operative treatment was administered. Additional details of the original tumor were not available for review.

CT with contrast of the current abdominal mass showed evidence of a heterogenous enhancing masses in the left upper abdominal quadrant. Additionally, there were multiple intrahepatic low attenuation lesions. Imaging interpretation suggested recurrent gastrointestinal stromal tumor with liver metastasis, and it was recommended for tissue confirmation. The largest mass in the liver measured 4 X 3 cm and the largest abdominal mass measured 6 x 5 cm, which was located at the pancreatic tail and spleen.

The pathological differential diagnosis was a recurrent gastrointestinal stromal tumor versus pancreatic neuroendocrine tumor or other types of mesenchymal tumors. A final needle aspiration biopsy (FNA) with adequate cellblock preparation

from one of the liver masses was obtained and the diagnosis was compatible with a metastatic high grade gastrointestinal stromal tumor.

The patient received neoadjuvant treatment utilizing the tyrosine kinase inhibitor Imatinib, followed by debulking surgical removal of the retroperitoneal abdominal masses. Surgical removal included the tumor masses, left adrenal gland, spleen, and portions of the pancreas and stomach. (Figure-1A). There was no attempt to remove liver masses. Microscopic examination showed histomorphologic features of mixed highly atypical spindle and epithelioid cells with prominent malignant giant tumor cells and abundant mitosis (more than 12 mitosis/10 HPF) (Figure 1-B-C). Vascular invasion was easily identified (Figure 1D).

Immunohistochemistry (IHC) studies were utilized for evaluation. The tumor cells were positive for Vimentin, CD34/QBEND-10 and CD117 (c-Kit) (Figure 1E). The tumor cells were negative for Cytokeratin AE1/AE3, S-100, SMA, Desmin, CD31, Pan Melanoma, CD-56, Synaptophysin, Chromogranin, and CDX-2.

The histomorphology together with the IHC profile and the patient's history was diagnostic of recurrent malignant gastrointestinal stromal tumor with multiple liver metastasis. The excised tumor mass showed more than 75% non-viable post-chemotherapy necrosis and fibrosis. Surgical margins were not free in multiple sites. Patient received another round of post-operative chemotherapy plus radiation and was free of recurrence of tumor for 9 months follow up, after which he expired due to massive metastatic diseases and multiple organ failure.

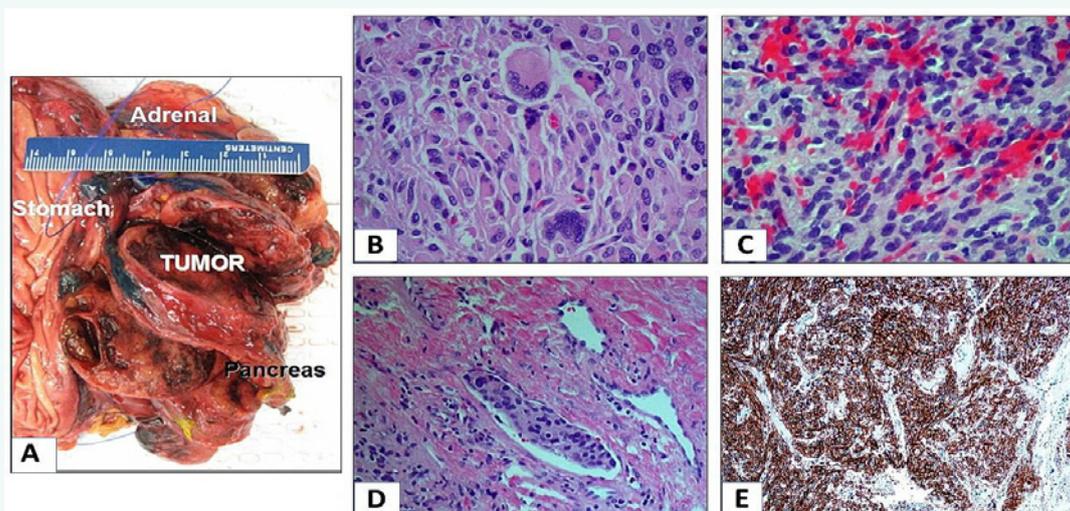


Figure 1 Pathological examination of the tumor

1A: Excised surgical mass including left adrenal, tumor, spleen, and portions of stomach and pancreas

1B: Epithelioid tumor cells with frequent multinucleation and increased mitosis cells (H&E stain X60)

1C: Spindle tumor cells with moderate atypia (H&E stain X40)

1D: Positive CD117



DISCUSSION

GISTs represent 80% of all gastrointestinal mesenchymal tumors. 60% of GISTs are found in the stomach and 20-30% are found in the small intestine, thought they can be found outside of the gastrointestinal tract via metastasis from a primary neoplasm [3,4]. GISTs arising outside of the gastrointestinal tract are thought to arise from interstitial cells of Cajal that were accidentally dispersed during the development of the embryo.¹⁶ Recently, investigators are suggesting that all GISTs are considered potentially malignant and are therefore classified based on rate of recurrence and risk of metastasis [12-14].

Clinically, most GISTs are asymptomatic and are discovered incidentally during an endoscopic procedure or on an imaging analysis done for another purpose. More often, they are associated with vague symptoms such as bloating, abdominal pain or discomfort, and early satiety; some may ulcerate, bleed, or grow large enough to cause gastrointestinal obstruction [17]. Consumptive hypothyroidism and non-islet cell tumor hypoglycemia are paraneoplastic syndromes associated with GISTs, though these are rare occurrences [18]. GISTs frequently metastasize to the peritoneum and liver, though rarely to regional lymph nodes or lungs, the most common site for soft tissue sarcoma metastasis [12]. GIST recurrence is determined based on classification systems developed by the National Institute of Health (NIH) consensus criteria and the Armed Forces Institute of Pathology criteria, which are based on mitotic count, tumor size, primary tumor site, and tumor rupture [19]. The recurrent tumor in our case was classified as malignant with high potential for recurrence and metastasis.

Radiologically, GIST usually appears as solid, smoothly contoured masses that enhance brightly with IV contrast [12]. Tumors larger than 15 cm may appear more compound due to hemorrhage, degeneration, and necrosis. Leiomyomas and GISTs may appear as submucosal masses with smooth margins, with an overlying mucosa and central ulceration.

Standard endoscopic biopsy techniques do not obtain sufficient tissue for diagnosis [20]. Snare biopsies are generally avoided for GISTs, as they can result in bowel perforation. Preoperative biopsy is not recommended for a resectable lesion in which there is a high degree of suspicion and radiological evidence of a GIST. However, biopsy is preferred to confirm metastatic disease or if preoperative tyrosine kinase inhibitors are considered prior to attempting resection in patients who have a large lesion considered to be GIST. The use of immunohistochemistry for KIT protein expression, polymerase chain reaction (PCR) for KIT mutations, and cytologic analysis may be used in diagnosing these lesions by endoscopic ultrasonography-guided fine-needle aspiration (EUS-FNA). In our case, diagnosis was confirmed by EUS-FNA of one of the metastatic liver lesions.

GISTs were considered rare tumors until 2000, often being misclassified as leiomyomas, leiomyoblastomas, or leiomyosarcomas [1]. The current understanding of GISTs is that they arise from interstitial cells of Cajal, which are derived from mesoderm and serve as the pacemaker cells for the

gastrointestinal tract [5,6]. Cajal cells characteristically stain positive for CD117, CD34, DOG-1 and protein kinase C theta [21-23]. The discovery of CD117 expression by GISTs was a scientific breakthrough in differentiating GISTs from leiomyosarcomas, leiomyomas, and other spindle cell tumors of the GI tract, which are CD117 negative [24]. CD117 antigen is compatible with the c-KIT receptor tyrosine kinase, a product of the KIT protooncogene. In normal cells, KIT receptor tyrosine kinase activity is regulated by binding of the KIT ligand or stem cell factor (SCF) to the endogenous ligand [25]. Mutations in KIT lead to constitutive activation of KIT, resulting in stimulation of cell survival, growth, and proliferation [26]. Numerous studies have indicated that approximately 80% of GISTs can carry a mutation for the KIT gene, allowing a structural variant of the KIT protein to be formed, which can be abnormally activated to enable oncogenic signaling [24]. KIT mutations in GIST can occur in different exons of the gene, with 70% affecting exon 11, which codes for the intracellular juxtamembrane domain of the receptor, and 12-15% affecting exon 9, which codes for the extracellular ligand-binding domain [24,26]. Activating mutations in the KIT gene are associated with GISTs, testicular seminomas, mast cell disease, melanoma, and acute myeloid leukemia [27]. Approximately 15% of GISTs lack mutations in KIT. A set of GISTs lacking KIT mutations have activating mutations in platelet-derived growth factor receptor alpha (PDGFRA) receptor tyrosine kinase [28-30].

GISTs represent a problematic subset of gastrointestinal neoplasms due to the variability of immunohistochemistry and cytogenetics of the tumors across patients. GISTs are mainly identified by expression of the KIT protein and activating mutations in the KIT or platelet-derived growth factor receptor alpha (PDGFRA) genes. Although the majority of GISTs are sporadic, 5% of patients diagnosed with primary familial GIST syndrome, neurofibromatosis type 1, and Carney-Stratakis syndrome carry a predisposition [9]. Individuals with neurofibromatosis I have a high incidence of GISTs in the small intestine and primarily do not carry somatic mutations in the KIT or PDGFRA genes. GISTs are rare in pediatric patients, but are linked to mutations in succinate dehydrogenase, as well as Carney-Stratakis syndrome [10,31]. Interestingly, the majority of pediatric GISTs have distinct characteristics from adult GISTs, including the absence of a mutation in C-kit tyrosine kinase or PDGFRA in 85% of cases as well as high incidence of lymph node metastases [10,11,32]. Moreover, most adult GISTs occur sporadically, while pediatric GISTs arise due to defined syndromes.

GISTs range in their cellular morphology, with the majority being spindle-cell type and the minority belonging to either the epithelioid-cell type or mixed type. Spindle-cell type GISTs are composed of uniform eosinophilic cells arranged in whorls or short fascicles [33]. When compared with leiomyomas, spindle-cell GISTs have paler eosinophilic cytoplasm and a fibrillary appearance. Epithelioid-type GISTs are composed of rounded cells with variably eosinophilic or clear cytoplasm with round to oval nuclei, vesicular, and nested architecture; possibly causing confusion with a melanocytic or epithelial neoplasm



[34]. However, epithelioid-type GISTs are often KIT-expression negative and have platelet derived growth factor receptor alpha (PDGFRA) mutations. Consequently, caution should be exercised when typing these neoplasms. The tumor in our case was of mixed morphology with strong positivity for CD-117 and CD34

There are no effective preventative measures for GIST, as it is linked with advanced age and certain inherited conditions. Early detection combined with complete resection of the mass, administration of tyrosine kinase inhibitors, as well as observation for possible recurrence is considered the current standard of care in patients with GIST [15]. Our presented case shows evidence of recurrence 2 years after initial GIST.

Prior to 2000, there was no effective therapy for metastatic or unresectable GISTs. Studies have shown that sarcomas of the GI tract have a low response rate to chemotherapy, proving that GISTs have a higher rate of primary resistance to chemotherapy [35-37]. The discovery of mutational activation of PDGFRA or KIT revolutionized treatments of GISTs, leading to the development of receptor tyrosine kinase inhibitors. Receptor tyrosine kinase inhibitors block signaling of PDGFRA or KIT by binding the ATP-binding pocket needed for phosphorylation to activate the receptor [38].

This case and literature review seeks to shed light on the variability, diagnostic and treatment challenges in patients who have GIST. Early detection combined with complete resection of the mass, administration of tyrosine kinase inhibitors, as well as observation for possible recurrence is considered the current standard of care in patients with GIST, however, most GISTs are found incidentally [15]. This delay in early detection poses an additional hurdle due to the potential malignancy and metastatic spread of most GISTs. The definitive diagnosis is made through immunohistochemical and radiological findings, which aids in the correct diagnosis of these neoplasms.

ACKNOWLEDGMENT

Special thanks to Iqra bhatti, Luna Emogene, and Sara Solomon, MD candidates, American University of the Caribbean for their assistance in reviewing the final manuscript

REFERENCES

1. Ashoor, A. A., & Barefah, G. (2020). Unusual presentation of a large GIST in an extraintestinal site: a challenging diagnosis dilemma. *BMJ case reports*, 13(2), e229839. <https://doi.org/10.1136/bcr-2019-229839>.
2. Hirota S. (2018). Differential diagnosis of gastrointestinal stromal tumor by histopathology and immunohistochemistry. *Translational gastroenterology and hepatology*, 3, 27. <https://doi.org/10.21037/tgh.2018.04.01>
3. LLENAS-GARCÍA J., GUERRA-VALES J. M., MORENO A., IBARROLA C., CASTELBON F. J., FERNÁNDEZ-RUIZ M., MENEU J. C., BALLESTIN C., MORENO E., Primary extragastrointestinal stromal tumors in the omentum and mesentery: a clinicopathological and immunohistochemical study, *Hepatogastroenterology*, 2008, 55(84):1002-1005.
4. Miettinen M, Monihan JM, Sarlomo-Rikala M, et al. Gastrointestinal stromal tumors/smooth muscle tumors (GISTs) primary in the omentum and mesentery: clinicopathologic and immunohistochemical study of 26 cases. *Am J Surg Pathol*. 1999;23:1109-1118.
5. Kindblom, L. G., Remotti, H. E., Aldenborg, F., & Meis-Kindblom, J. M. (1998). Gastrointestinal pacemaker cell tumor (GIPACT): gastrointestinal stromal tumors show phenotypic characteristics of the interstitial cells of Cajal. *The American journal of pathology*, 152(5), 1259-1269.
6. Kursad Turksen (2006). *Embryonic Stem Cell Protocols: Differentiation models*. Humana Press. pp. 263-. ISBN 978-1-58829-784-6. Retrieved 14 April 2010.
7. Miettinen M, Lasota J. Gastrointestinal stromal tumors--definition, clinical, histological, immunohistochemical, and molecular genetic features and differential diagnosis. *Virchows Arch*. 2001 Jan;438(1):1-12. doi: 10.1007/s004280000338. PMID: 11213830.
8. Ma GL, Murphy JD, Martinez ME, Sicklick JK. Epidemiology of gastrointestinal stromal tumors in the era of histology codes: results of a population-based study. *Cancer Epidemiol Biomarkers Prev*. 2015 Jan;24(1):298-302. doi: 10.1158/1055-9965.EPI-14-1002. Epub 2014 Oct 2. PMID: 25277795; PMCID: PMC4294949.
9. Mussi C, Schildhaus HU, Gronchi A, Wardelmann E, Hohenberger P. Therapeutic consequences from molecular biology for gastrointestinal stromal tumor patients affected by neurofibromatosis type 1. *Clin Cancer Res*. 2008 Jul 15;14(14):4550-5. doi: 10.1158/1078-0432.CCR-08-0086. PMID: 18628470.
10. Janeway KA, Pappo A. Treatment guidelines for gastrointestinal stromal tumors in children and young adults. *J Pediatr Hematol Oncol*. 2012 May;34 Suppl 2:S69-72. doi: 10.1097/MPH.0b013e31824e3899. PMID: 22525410.
11. Pappo AS, Janeway KA. Pediatric gastrointestinal stromal tumors. *Hematol Oncol Clin North Am*. 2009 Feb;23(1):15-34, vii. doi: 10.1016/j.hoc.2008.11.005. PMID: 19248968.
12. Morgan J, Raut CP, Duensing A, Keedy VL. Epidemiology, classification, clinical presentation, prognostic features, and diagnostic work-up of gastrointestinal stromal tumors (GIST). UpToDate. 2019. Accessed at <https://www.uptodate.com/contents/epidemiology-classification-clinical-presentation-prognostic-features-and-diagnostic-work-up-of-gastrointestinal-stromal-tumors-gist> on January 21, 2021.
13. Miettinen M, Lasota J. Gastrointestinal stromal tumors: pathology and prognosis at different sites. *Semin Diagn Pathol*. 2006 May;23(2):70-83. doi: 10.1053/j.semmp.2006.09.001. PMID: 17193820.
14. Huang HY, Li CF, Huang WW, Hu TH, Lin CN, Uen YH, Hsiung CY, Lu D. A modification of NIH consensus criteria to better distinguish the highly lethal subset of primary localized gastrointestinal stromal tumors: a subdivision of the original high-risk group on the basis of outcome. *Surgery*. 2007 Jun;141(6):748-56. doi: 10.1016/j.surg.2007.01.024. Epub 2007 May 4. PMID: 17560251.
15. Vallilas C, Sarantis P, Kyriazoglou A, Koustas E, Theocharis S, Papavassiliou AG, Karamouzis MV. Gastrointestinal Stromal Tumors (GISTs): Novel Therapeutic Strategies with Immunotherapy and Small Molecules. *Int J Mol Sci*. 2021 Jan 6;22(2):493. doi: 10.3390/ijms22020493. PMID: 33419029; PMCID: PMC7825300.
16. Wang L, Vargas H, French SW. Cellular origin of gastrointestinal stromal tumors: a study of 27 cases. *Arch Pathol Lab Med*. 2000 Oct;124(10):1471-5. doi: 10.1043/0003-9985(2000)124<1471:COO GST>2.0.CO;2. PMID: 11035578.
17. Miettinen M, Makhlof H, Sobin LH, Lasota J. Gastrointestinal stromal tumors of the jejunum and ileum: a clinicopathologic,



- immunohistochemical, and molecular genetic study of 906 cases before imatinib with long-term follow-up. *Am J Surg Pathol.* 2006 Apr;30(4):477-89. doi: 10.1097/00000478-200604000-00008. PMID: 16625094.
18. Maynard MA, Marino-Enriquez A, Fletcher JA, Dorfman DM, Raut CP, Yassa L, Guo C, Wang Y, Dorfman C, Feldman HA, Frates MC, Song H, Jugo RH, Taguchi T, Hershman JM, Larsen PR, Huang SA. Thyroid hormone inactivation in gastrointestinal stromal tumors. *N Engl J Med.* 2014 Apr 3;370(14):1327-34. doi: 10.1056/NEJMoa1308893. PMID: 24693892; PMCID: PMC4186889.
19. Jang SH, Kwon JE, Kim JH, Lee JY, Kim SG, Kim JS, Jung HC, Im JP. Prediction of Tumor Recurrence in Patients with Non-Gastric Gastrointestinal Stromal Tumors Following Resection according to the Modified National Institutes of Health Criteria. *Intest Res.* 2014 Jul;12(3):229-35. doi: 10.5217/ir.2014.12.3.229. Epub 2014 Jul 25. PMID: 25349597; PMCID: PMC4204716.
20. Watson RR, Binmoeller KF, Hamerski CM, Shergill AK, Shaw RE, Jaffee IM, Stewart L, Shah JN. Yield and performance characteristics of endoscopic ultrasound-guided fine needle aspiration for diagnosing upper GI tract stromal tumors. *Dig Dis Sci.* 2011 Jun;56(6):1757-62. doi: 10.1007/s10620-011-1646-6. Epub 2011 Mar 1. PMID: 21360279.
21. Novelli M, Rossi S, Rodriguez-Justo M, Taniere P, Seddon B, Toffolatti L, Sartor C, Hogendoorn PC, Sciot R, Van Glabbeke M, Verweij J, Blay JY, Hohenberger P, Flanagan A, Dei Tos AP. DOG1 and CD117 are the antibodies of choice in the diagnosis of gastrointestinal stromal tumours. *Histopathology.* 2010 Aug;57(2):259-70. doi: 10.1111/j.1365-2559.2010.03624.x. PMID: 20716168.
22. Miettinen M, Wang ZF, Lasota J. DOG1 antibody in the differential diagnosis of gastrointestinal stromal tumors: a study of 1840 cases. *Am J Surg Pathol.* 2009 Sep;33(9):1401-8. doi: 10.1097/PAS.0b013e3181a90e1a. PMID: 19606013.
23. Kang GH, Srivastava A, Kim YE, Park HJ, Park CK, Sohn TS, Kim S, Kang DY, Kim KM. DOG1 and PKC- θ are useful in the diagnosis of KIT-negative gastrointestinal stromal tumors. *Mod Pathol.* 2011 Jun;24(6):866-75. doi: 10.1038/modpathol.2011.11. Epub 2011 Feb 25. PMID: 21358619.
24. Hirota S, Isozaki K, Moriyama Y, Hashimoto K, Nishida T, Ishiguro S, Kawano K, Hanada M, Kurata A, Takeda M, Muhammad Tunio G, Matsuzawa Y, Kanakura Y, Shinomura Y, Kitamura Y. Gain-of-function mutations of c-kit in human gastrointestinal stromal tumors. *Science.* 1998 Jan 23;279(5350):577-80. doi: 10.1126/science.279.5350.577. PMID: 9438854
25. Broudy VC. Stem cell factor and hematopoiesis. *Blood.* 1997 Aug 15;90(4):1345-64. PMID: 9269751.
26. Corless CL, Barnett CM, Heinrich MC. Gastrointestinal stromal tumours: origin and molecular oncology. *Nat Rev Cancer.* 2011 Nov 17;11(12):865-78. doi: 10.1038/nrc3143. PMID: 22089421.
27. KIT KIT proto-oncogene, receptor tyrosine kinase [Homo sapiens (human)] - Gene - NCBI. (n.d.). Retrieved January 29, 2021, from <https://www.ncbi.nlm.nih.gov/gene?Db=gene&Cmd=ShowDetailView&TermToSearch=3815>
28. Medeiros F, Corless CL, Duensing A, Hornick JL, Oliveira AM, Heinrich MC, Fletcher JA, Fletcher CD. KIT-negative gastrointestinal stromal tumors: proof of concept and therapeutic implications. *Am J Surg Pathol.* 2004 Jul;28(7):889-94. doi: 10.1097/00000478-200407000-00007. PMID: 15223958.
29. Hirota S, Ohashi A, Nishida T, Isozaki K, Kinoshita K, Shinomura Y, Kitamura Y. Gain-of-function mutations of platelet-derived growth factor receptor alpha gene in gastrointestinal stromal tumors. *Gastroenterology.* 2003 Sep;125(3):660-7. doi: 10.1016/s0016-5085(03)01046-1. PMID: 12949711.
30. Martín J, Poveda A, Llombart-Bosch A, Ramos R, López-Guerrero JA, García del Muro J, Maurel J, Calabuig S, Gutierrez A, González de Sande JL, Martínez J, De Juan A, Láinez N, Losa F, Alija V, Escudero P, Casado A, García P, Blanco R, Buesa JM; Spanish Group for Sarcoma Research. Deletions affecting codons 557-558 of the c-KIT gene indicate a poor prognosis in patients with completely resected gastrointestinal stromal tumors: a study by the Spanish Group for Sarcoma Research (GEIS). *J Clin Oncol.* 2005 Sep 1;23(25):6190-8. doi: 10.1200/JCO.2005.19.554. Erratum in: *J Clin Oncol.* 2006 Apr 10;24(11):1784. García, Pilar [corrected to García, Paula]. PMID: 16135486.
31. Janeway KA, Kim SY, Lodish M, Nosé V, Rustin P, Gaal J, Dahia PL, Liegl B, Ball ER, Raygada M, Lai AH, Kelly L, Hornick JL; NIH Pediatric and Wild-Type GIST Clinic, O'Sullivan M, de Krijger RR, Dinjens WN, Demetri GD, Antonescu CR, Fletcher JA, Helman L, Stratakis CA. Defects in succinate dehydrogenase in gastrointestinal stromal tumors lacking KIT and PDGFRA mutations. *Proc Natl Acad Sci U S A.* 2011 Jan 4;108(1):314-8. doi: 10.1073/pnas.1009199108. Epub 2010 Dec 20. PMID: 21173220; PMCID: PMC3017134.
32. Agaram NP, Laquaglia MP, Ustun B, Guo T, Wong GC, Socci ND, Maki RG, DeMatteo RP, Besmer P, Antonescu CR. Molecular characterization of pediatric gastrointestinal stromal tumors. *Clin Cancer Res.* 2008 May 15;14(10):3204-15. doi: 10.1158/1078-0432.CCR-07-1984. PMID: 18483389; PMCID: PMC3805121.
33. Fletcher CD, Berman JJ, Corless C, Gorstein F, Lasota J, Longley BJ, Miettinen M, O'Leary TJ, Remotti H, Rubin BP, Shmookler B, Sobin LH, Weiss SW. Diagnosis of gastrointestinal stromal tumors: a consensus approach. *Int J Surg Pathol.* 2002 Apr;10(2):81-9. doi: 10.1177/106689690201000201. PMID: 12075401.
34. Miettinen M, Sobin LH, Sarlomo-Rikala M. Immunohistochemical spectrum of GISTs at different sites and their differential diagnosis with a reference to CD117 (KIT). *Mod Pathol.* 2000 Oct;13(10):1134-42. doi: 10.1038/modpathol.3880210. PMID: 11048809.
35. Demetri GD, Elias AD. Results of single-agent and combination chemotherapy for advanced soft tissue sarcomas. Implications for decision making in the clinic. *Hematol Oncol Clin North Am.* 1995 Aug;9(4):765-85. PMID: 7490240.
36. Plaat BE, Hollema H, Molenaar WM, Torn Broers GH, Pijpe J, Mastik MF, Hoekstra HJ, van den Berg E, Scheper RJ, van der Graaf WT. Soft tissue leiomyosarcomas and malignant gastrointestinal stromal tumors: differences in clinical outcome and expression of multidrug resistance proteins. *J Clin Oncol.* 2000 Sep 15;18(18):3211-20. doi: 10.1200/JCO.2000.18.18.3211. PMID: 10986053.
37. Edmonson JH, Marks RS, Buckner JC, Mahoney MR. Contrast of response to dacarbazine, mitomycin, doxorubicin, and cisplatin (DMAP) plus GM-CSF between patients with advanced malignant gastrointestinal stromal tumors and patients with other advanced leiomyosarcomas. *Cancer Invest.* 2002;20(5-6):605-12. doi: 10.1081/cnv-120002485. PMID: 12197215.
38. Balachandran VP, Cavnar MJ, Zeng S, Bamboat ZM, Ocuin LM, Obaid H, Sorenson EC, Popow R, Ariyan C, Rossi F, Besmer P, Guo T, Antonescu CR, Taguchi T, Yuan J, Wolchok JD, Allison JP, DeMatteo RP. Imatinib potentiates antitumor T cell responses in gastrointestinal stromal tumor through the inhibition of Ido. *Nat Med.* 2011 Aug 28;17(9):1094-100. doi: 10.1038/nm.2438. PMID: 21873989; PMCID: PMC3278279.